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References:

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2. JCVI: [MT3000](#)
3. TubercuList: Gene [Rv2931](#)
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SOP: TN002
Obtaining Cells from Inoculated Transposon Mutant LJ Slants
Materials and reagents:

1. *M. tuberculosis*, transposon mutant LJ slant
2. Biosafety cabinet
3. Sterile aerosol resistant pipet tips, 200 µL
4. Pipetman, 200 µL
5. Cell scraper, sterile
6. 7H9 media (note 3)
7. 7H11 + OADC agar plate, 100 x 15 mm (VWR catalog no. 29447-102)
8. Cold room or 4°C refrigerator

Protocol:

1. Remove LJ slant from container within biosafety cabinet (note 1).
2. Add 200 µL of 7H9 media to LJ slant.
3. Use cell scraper to lightly scrape the cells on the LJ slant into the 7H9 media.
4. Pipet 100 µL of the media, which now contains cell growth, onto a small 7H11 + OADC plate (note 2).
5. Streak the bacteria to grow as a lawn.
6. Place inoculated plates in a Ziploc bag, seal, and place in warm room (note 4).
7. Once cells have grown, move plates into biosafety cabinet (note 5).
8. Inside the biosafety cabinet, use a sterile cell scraper and aseptically scrape the cells into GAS media or liquid media of choice.

Notes:

1. The LJ slants must be removed from packaging only within a BSL3 facility and opened only within a BSL3 biosafety cabinet.
2. Use an aerosol resistant tip and pipetman to transfer cells from the liquid culture to the 7H11 plate. If preparing your own agar plates, follow the instructions on the bottle of 7H11 powder (Fisher Scientific catalog # DF0838-17-9).
3. Follow the instructions on the bottle of 7H9 powder (VWR catalog # 90003-876).
4. LJ slants can be kept in a cold room or 4°C refrigerator for future use.
5. Depending upon the strain, a lawn could take 2 to 4 weeks to form.