

Respiratory Syncytial Virus (RSV) A2 Nucleoprotein (N) Helper Plasmid, pA2-Nopt

Catalog No. NR-36462

For research use only. Not for human use.

Contributor:

BEI Resources

Manufacturer:

Martin L. Moore, Assistant Professor, Department of Pediatrics, Division of Infectious Diseases, Emory University School of Medicine, Atlanta, Georgia, USA

Product Description:

NR-36462 is a component of a bacterial artificial chromosome (BAC)-based RSV rescue system that allows RSV infection to be monitored by fluorescence and is an important tool in RSV vaccine research and mutagenesis studies. Please refer to Appendix I for the manufacturer's RSV rescue protocol.

The N helper plasmid was constructed from codon-optimized RSV A2 N sequences. The codon-optimized cDNA sequences were synthesized and cloned into the pcDNA™3.1(+)⁴ mammalian expression plasmid (Life Technologies™ Invitrogen™).^{1,2} The plasmid was produced in *Escherichia coli*, strain 10-beta (a DH10B derivative, New England BioLabs®) and extracted using a Endo-Free Plasmid Maxi Kit (Qiagen).² The complete sequence for pA2-Nopt is reported in Appendix II.

Material Provided:

Each vial contains 0.5 µg of plasmid DNA in RNase/DNase-free 10 mM Tris-HCl, 1 mM EDTA buffer (pH 8). The concentration is shown on the Certificate of Analysis. The vial should be centrifuged prior to opening.

Packaging/Storage:

NR-36462 was packaged aseptically in screw-capped plastic cryovials. The product is provided frozen on dry ice and should be stored at -80°C or colder immediately upon arrival. Freeze-thaw cycles should be minimized.

Functional Activity:

Recombinant RSV was produced by co-transfection of BHK-21 clone BSR T7/5 cells³ with pSynkRSV-I19F, a BAC plasmid containing RSV A2-line19F antigenomic DNA and the gene for the far-red fluorescent protein monomeric Katushka 2 (mKate2) to enable detection of infection through fluorescence, (NR-36460) and four helper plasmids encoding sequence-optimized genes from RSV strain A2: large polymerase (L) (NR-36461), nucleoprotein (N) (NR-36462), phosphoprotein (P) (NR-36463) and matrix 2-1 protein (M2-1) (NR-36464). RSV rescue and infection could be detected by red fluorescent syncytia.

Citation:

Acknowledgment for publications should read "The following reagent was obtained through BEI Resources, NIAID, NIH: Respiratory Syncytial Virus (RSV) A2 Nucleoprotein (N) Helper Plasmid, pA2-Nopt, NR-36462."

Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the following publication: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, and National Institutes of Health. Biosafety in Microbiological and Biomedical Laboratories, 5th ed. Washington, DC: U.S. Government Printing Office, 2009; see www.cdc.gov/biosafety/publications/bmb15/index.htm.

Disclaimers:

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References:

- Hotard, A. L., et al. "A Stabilized Respiratory Syncytial Virus Reverse Genetics System Amendable to Recombination-Mediated Mutagenesis." Virology 434 (2012): 129-136. PubMed: 23062737.

2. M. L. Moore, Personnel Communication.
3. Buchholz, U. J., S. Finke and K. -K. Conzelmann. "Generation of Bovine Respiratory Syncytial Virus (BRSV) from cDNA: BRSV NS2 Is Not Essential for Virus Replication in Tissue Culture, and Human RSV Leader Region Acts as a Functional BRSV Genome Promoter." J. Virol. 73 (1999): 251-259. PubMed: 9847328.

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Appendix I

Transfection Procedure for Virus Recovery of Recombinant Respiratory Syncytial Virus

Materials (Suggested suppliers and catalog numbers are indicated):

BHK-21 clone BSR T7/5 cell cultures or alternative cells [BHK21 cells (ATCC® CCL10™) transfected with phage T7 polymerase from Modified Vaccinia Ankara (MVA)] **Note:** This protocol is optimized for use with BHK-21 clone BSR T7/5 cells. Use of alternative cells may result in decreased recovery of RSV.

Opti-MEM (serum-free) (Gibco/Life Technologies catalog #11058-021)

GMEM [Glasgow's MEM (Gibco/Life Technologies catalog #11710-035)] + 3% FBS

MEM non-essential amino acids (NEAA) 100X solution (Gibco/Life Technologies catalog #11140-050)

G418 sulfate, 50 mg/mL solution (500X) (Agilent Technologies Genomics catalog # 200049)

Trypsin-EDTA (0.25%) (Gibco/Life Technologies catalog #25200-072)

Antibiotic-Antimycotic solution, penicillin/streptomycin/amphotericin (100X) (Corning cellgro® catalog #30-004-CI) or equivalent

Plasmid with RSV antigenome (NR-36460) each vial contains 0.5 µg in 5 µL total volume **Note:** This protocol requires 0.8 µg of pSynkRSV-I19F; thus 2 vials of NR-36460 are required per transfection.)

Helper Plasmids – (all codon optimized) each vial contains 0.5 µg in 5 µL total volume:

pA2-Lopt, L protein (NR-36461)

pA2-Nopt, N protein (NR-36462)

pA2-Popt, P protein (NR-36463)

pA2-M2-1opt, M2-1 protein (NR-36464)

Lipofectamine 2000 transfection reagent (Gibco/Life Technologies catalog #11668-019)

Phosphate buffered saline pH 7.2 (Gibco/Life Technologies catalog #20012027)

6-well tissue culture plates

25 cm² tissue culture flasks

Shaker/rocker plate

Tissue culture humidified incubator with 3% to 5% CO₂

Assorted sterile pipettes and tips

Procedure:

Note: This protocol assumes the user is familiar with cell culture techniques and transfection procedures.

1. Initial cell culture:
 - a. For routine sub-passage of BSR T7/5 cells, prepare new 25 cm² cultures at a ratio of one donor culture to three new cultures, based on surface area of the culture flasks (1:3 passage ratio). Use GMEM with 3% FBS + 1X NEAA + 1X antibiotics as growth medium, 5 mL per flask. When maintaining donor cultures, add 1X G418 to the growth medium every other passage.
 - b. For transfections, sub-pass BSR T7/5 cells from “donor” cultures into 6 well plates so they will be 100% confluent at time of transfection. Use one 25 cm² culture to prepare one 6 well plate (1:2.5 passage ratio).
2. Prepare 6 well plates for transfection from 25 cm² donor cultures. Determine how many plates will be required and use the corresponding number of flasks. Aspirate the growth medium from the flasks, and then add 0.25 mL of warm trypsin-EDTA per 25 cm² flask. Rock flasks to distribute the trypsin-EDTA and incubate at 37°C for 5 to 10 minutes. When cells start to dislodge from the flask, add 12 mL of GMEM with 3% FBS to each flask and use a pipet to suspend the cells in this growth medium. Add 2 mL of the cell suspension to each well in the 6 well plates. Incubate the plates at 37°C in the tissue culture incubator until the cell sheets are confluent and ready for transfection.
3. Prepare the reagents for the transfection procedure. Transfection will be done using Lipofectamine 2000 as the transfection reagent. Additionally, it is important to include control transfections (Lipofectamine only/wild type virus for mutants etc.)
 - a. Use a 3:1 ratio of Lipofectamine (µL) to plasmid/helper plasmid (µg). Dilute each component with Opti-MEM to make 100 µL of each. After dilution, allow each dilution to sit at room temperature for 5 minutes.
 - b. Use the following amounts of each component per transfection:

i.	RSV antigenome (NR-36460)	0.8 µg (8 µL of 0.1 µg/µL) + 92 µL Opti-MEM
		(2 vials of NR-36460 are required per transfection.)
ii.	pA2-Lopt, L protein (NR-36461)	0.2 µg (2 µL of 0.1 µg/µL) + 98 µL Opti-MEM
iii.	pA2-Nopt, N protein (NR-36462)	0.4 µg (4 µL of 0.1 µg/µL) + 96 µL Opti-MEM
iv.	pA2-Popt, P protein (NR-36463)	0.4 µg (4 µL of 0.1 µg/µL) + 96 µL Opti-MEM
v.	pA2-M2-1opt, M2-1 protein (NR-36464)	0.4 µg (4 µL of 0.1 µg/µL) + 96 µL Opti-MEM

vi. Lipofectamine 2000

6.6 μ L + 93.4 μ L Opti-MEM

Note: For multiple transfections increase the above quantities proportionally.

- c. After allowing the diluted components to sit at room temperature for 5 minutes, combine all six components in one vial, mix gently and incubate the transfection mixture at room temperature for 20 minutes.
 - d. Transfection mixtures should be 600 μ L total (Opti-MEM, Lipofectin, and DNA)
 - e. Aspirate the media from the BSR T7/5 cell culture plate, wash cells twice with 1 mL warm Opti-MEM for each wash, and aspirate the final wash.
 - f. Add 600 μ L transfection mixture to each well and incubate the plate 2 hours at room temperature on a shaker/rocker plate set at low speed.
 - g. After 2 hours, add an additional 600 μ L warm Opti-MEM per well and place plate in a 37°C tissue culture incubator overnight (8-12 hours).
4. After incubation, aspirate and discard the transfection mixture from the wells, wash each well once with 1 mL warm sterile PBS, aspirate the PBS and replace with 2 mL of warm GMEM with 3% FBS per well. Continue incubating at 37°C in the tissue culture incubator overnight.
 5. Day 2 post transfection, sub-pass the cells into 25 cm² flasks using the trypsin-EDTA procedure described above. Pass at a 1:3 surface area ratio unless cell morphology appears weak, in which case the ratio should be decreased accordingly up to an even 1:1 ratio. (Note: surface area of each well in the 6 well plate is 10 cm²). Cells should remain in GMEM with 3% FBS throughout the rest of recovery.
 6. Monitor flasks for cytopathic effect (CPE) and sub-pass at 1:3 ratio into new 25 cm² flasks as needed (approximately every 48 hours). CPE shows first as mini-syncytia and then grows into rounded up clumps of cells.
 7. When CPE is evident throughout the flask, scrape the cells into the growth media and aliquot into cryovials. Freeze at -80°C or colder.

Appendix 2: pA2-Nopt Sequence

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1   GACGGATCGGGAGATCTCCCGATCCCCTATGGTGCACCTCTCAGTACAATCTGCTCTGATG 60
   CTGCCTAGCCCTCTAGAGGGGCTAGGGGATACCACGTGAGAGTCATGTTAGACGAGACTAC

61   CCGCATAGTTAAGCCAGTATCTGCTCCCTGCTTGTGTGTTGGAGGTCGCTGAGTAGTGCG 120
   GCGGTATCAATTCCGGTCATAGACGAGGGACGAACACACAACCTCCAGCGACTCATCACGC

121  CGAGCAAAATTTAAGCTACAACAAGGCAAGGCTTGACCGACAATTGCATGAAGAATCTGC 180
   GCTCGTTTTAAATTCGATGTTGTTCCGTTCCGAACCTGGCTGTTAACGTACTTCTTAGACG

181  TTAGGGTTAGGCGTTTTGCGCTGCTTCGCGATGTACGGGCCAGATATACGCGTTGACATT 240
   AATCCCAATCCGCAAAACGCGACGAAGCGCTACATGCCCGGTCTATATGCGCAACTGTAA

241  GATTATTGACTAGTTATTAATAGTAATCAATTACGGGGTCATTAGTTCATAGCCCATATA 300
   CTAATAACTGATCAATAATTATCATTAGTTAATGCCCCAGTAATCAAGTATCGGGTATAT

301  TGGAGTTCGCGGTTACATAACTTACGGTAAATGGCCCGCCTGGCTGACCGCCCAACGACC 360
   ACCTCAAGGCGCAATGTATTGAATGCCATTTACGGGGCGGACCGACTGGCGGGTTGCTGG

361  CCCGCCCATGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTTCC 420
   GGGCGGGTAACTGCAGTTATTACTGCATACAAGGGTATCATTGCGGTTATCCCTGAAAGG

421  ATTGACGTCAATGGGTGGAGTATTTACGGTAAACTGCCCACTTGGCAGTACATCAAGTGT 480
   TAACTGCAGTTACCCACCTCATAAATGCCATTTGACGGGTGAACCGTCATGTAGTTCACA

481  ATCATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCCCGCCTGGCATT 540
   TAGTATACGGTTCATGCGGGGATAACTGCAGTTACTGCCATTTACGGGGCGGACCGTAA

541  ATGCCCAGTACATGACCTTATGGGACTTTCCTACTTGGCAGTACATCTACGTATTAGTCA 600
   TACGGGTCATGTACTGGAATACCCTGAAAGGATGAACCGTCATGTAGATGCATAATCAGT

601  TCGCTATTACCATGGTGATGCGGTTTTGGCAGTACATCAATGGGCGTGGATAGCGGTTTG 660
   AGCGATAATGGTACCCTACGCCAAAACCGTCATGTAGTTACCCGCACCTATCGCCAAAC

661  ACTCACGGGGATTTCCAAGTCTCCACCCATTGACGTCAATGGGAGTTTGTGTTTGGCACC 720
   TGAGTGCCCTAAAGGTTTCAGAGGTGGGGTAACTGCAGTTACCCTCAAACAAAACCGTGG

721  AAAATCAACGGGACTTTCCAAAATGTCGTAACAACCTCCGCCCCATTGACGCAAATGGGCG 780
   TTTTAGTTGCCCTGAAAGGTTTTACAGCATTGTTGAGGCGGGGTAACCTGCGTTTACCCGC

781  GTAGGCGTGACGGTGGGAGGTCTATATAAGCAGAGCTCTCTGGCTAACTAGAGAACCCA 840
   CATCCGCACATGCCACCCTCCAGATATATTCGTCTCGAGAGACCGATTGATCTCTTGGGT
           T7 promoter (863,881)
           |
841  CTGCTTACTGGCTTATCGAAATTAATACGACTCACTATAGGGAGACCCAAGCTGGCTAGC 900
   GACGAATGACCGAATAGCTTTAATTATGCTGAGTGATATCCCTCTGGGTTTCGACCGATCG
           KpnI      RSV nucleoprotein (929,2104)
           |      |
901  GTTTAAACTTAAGCTTGGTACCGCCACCATGGCCCTGAGCAAAGTGAAGCTGAACGACAC 960
   CAAATTTGAATTCGAACCATGGCGGTGGTACCGGGACTCGTTTCACTTCGACTTGCTGTG

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961 CCTGAACAAGGATCAGCTGCTGAGCAGCAGCAAGTACACCATCCAGCGGAGCACCGGCGA 1020
GGACTTGTTCCTAGTCGACGACTCGTCGTCGTTTCATGTGGTAGGTCGCCTCGTGGCCGCT

1021 CAGCATCGACACCCCCAACTACGACGTGCAGAAGCACATCAACAAGCTGTGCGGCATGCT 1080
GTCGTAGCTGTGGGGGTGATGCTGCACGTCTTCGTGTAGTTGTTTCGACACGCCGTACGA

1081 GCTGATCACCGAGGACGCCAACCACAAGTTCACCGGCCTGATCGGCATGCTGTACGCCAT 1140
CGACTAGTGGCTCCTGCGGTTGGTGTCAAGTGGCCGGACTAGCCGTACGACATGCGGTA

1141 GAGCAGACTGGGACGCGAGGACACCATCAAGATCCTGCGGGACGCCGGCTACCACGTGAA 1200
CTCGTCTGACCCTGCGCTCCTGTGGTAGTTCTAGGACGCCCTGCGGCCGATGGTGCACCTT

1201 GGCCAAATGGCGTGGACGTGACCACCCACCGGCAGGACATCAACGGCAAAGAAATGAAGTT 1260
CCGGTTACCGCACCTGCACTGGTGGGTGGCCGCTCCTGTAGTTGCCGTTTCTTTACTTCAA

1261 CGAGGTGCTGACCCTGGCCAGCCTGACCACCGAGATCCAGATCAACATCGAGATCGAGAG 1320
GCTCCACGACTGGGACCGGTGCGACTGGTGGCTCTAGGTCTAGTTGTAGCTCTAGCTCTC

1321 CCGGAAGTCCTACAAGAAAATGCTGAAAGAAATGGGCGAGGTGGCCCCGAGTACCGGCA 1380
GGCCTTCAGGATGTTCTTTTACGACTTTCTTTACCCGCTCCACCGGGGGCTCATGGCCGT

1381 CGATAGCCCCGACTGCGGCATGATCATCCTGTGTATCGCTGCCCTGGTGATCACAAAGCT 1440
GCTATCGGGGCTGACGCCGTACTAGTAGGACACATAGCGACGGGACCACTAGTGTTCGA

1441 GGCCGCTGGCGACAGATCCGGACTGACCGCTGTGATCAGACGGGCCAACAACGTGCTGAA 1500
CCGGCGACCGCTGTCTAGGCCTGACTGGCGACACTAGTCTGCCCGGTTGTTGCACGACTT

1501 GAACGAGATGAAGCGGTACAAGGGCCTGCTGCCAAGGATATCGCCAACAGCTTCTACGA 1560
CTTGCTCTACTTCGCCATGTTCCCGGACGACGGGTTCTATAGCGGTTGTGCAAGATGCT

1561 GGTGTTTCGAGAAGCACCCCCACTTCATCGACGTGTTTCGTGCACTTCGGAATCGCCCAGAG 1620
CCACAAGCTCTTCGTGGGGGTGAAGTAGCTGCACAAGCACGTGAAGCCTTAGCGGGTCTC

1621 CAGCACCAGAGGCGGCAGCCGGGTGGAAGGCATCTTCGCCGGCCTGTTTCATGAACGCCTA 1680
GTCGTGGTCTCCGCCGTCGGCCACCTTCCGTAGAAGCGGCCGGACAAGTACTTGCGGAT

1681 CGGCGCTGGCCAGGTGATGCTGAGATGGGGCGTGCTGGCCAAGAGCGTGAAGAACATCAT 1740
GCCGCGACCGGTCCACTACGACTCTACCCCGCACGACCGGTTCTCGCACTTCTTGTAGTA

1741 GCTGGGCCACGCCAGCGTGCAGGCCGAGATGGAACAGGTGGTGGAAAGTGTACGAGTACGC 1800
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1801 CCAGAAGCTGGGCGGCGAGGCCGGCTTCTACCACATCCTGAACAACCCCAAGGCCAGCCT 1860
GGTCTTCGACCCGCCGCTCCGGCCGAAGATGGTGTAGGACTTGTGGGGTTCCGGTCCGA

1861 GCTGTCCCTGACCCAGTTCCCCCACTTCTCCAGCGTGGTGGTGGCAATGCCGCCGACT 1920
CGACAGGACTGGGTCAAGGGGGTGAAGAGGTGCGACCACGACCCGTTACGGCGGCCTGA

1921 GGGCATCATGGGCGAGTACAGAGGCACCCCCCGGAACCAGGACCTGTACGACGCCCCAA 1980
CCCGTAGTACCCGCTCATGTCTCCGTGGGGGGCCTTGGTCTGGACATGCTGCGGCGGTT

1981 GGCCTACGCCGAGCAGCTGAAAGAAAACGGCGTGATCAACTACAGCGTGCTGGATCTGAC 2040
CCGGATGCGGCTCGTCGACTTTCTTTTGCCGCACTAGTTGATGTGCGCACGACCTAGACTG

2041 CGCCGAGGAACTGGAAGCCATCAAGCACCAGCTGAACCCCAAGGACAACGACGTGGAACT 2100
 GCGGCTCCTTGACCTTCGGTAGTTTCGTGGTGCAGCTTGGGGTTCCTGTTGCTGCACCTTGA
 XhoI
 |
 2101 GTGACTCGAGTCTAGAGGGCCCGTTTAAACCCGCTGATCAGCCTCGACTGTGCCTTCTAG 2160
 CACTGAGCTCAGATCTCCCGGGCAAATTTGGGCGACTAGTCGGAGCTGACACGGAAGATC

2161 TTGCCAGCCATCTGTTGTTTGGCCCTCCCCCGTGCCTTCCTTGACCCTGGAAGGTGCCAC 2220
 AACGGTCGGTAGACAACAAACGGGGAGGGGGCACGGAAGGAACTGGGACCTTCCACGGTG

2221 TCCCCTGTCCTTTTCTAATAAAAATGAGGAAATTGCATCGCATTGTCTGAGTAGGTGTCA 2280
 AGGGTGACAGGAAAGGATTATTTTACTCCTTTAACGTAGCGTAACAGACTCATCCACAGT

2281 TTCTATTCTGGGGGGTGGGGTGGGGCAGGACAGCAAGGGGGAGGATTGGGAAGACAATAG 2340
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2341 CAGGCATGCTGGGGATGCGGTGGGCTCTATGGCTTCTGAGGCGGAAAGAACCAGCTGGGG 2400
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 f1 origin(2435,2741)
 |
 2401 CTCTAGGGGGTATCCCCACGCGCCCTGTAGCGGCGCATTAAGCGCGGCGGGTGTGGTGGT 2460
 GAGATCCCCCATAGGGGTGCGCGGGACATCGCCGCGTAATTCGCGCCGCCACACCACCA

2461 TACGCGCAGCGTGACCGCTACACTTGCCAGCGCCCTAGCGCCCGCTCCTTTCGCTTTCTT 2520
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2521 CCCTTCCTTTCTCGCCACGTTTCGCCGGCTTTCCCCGTCAAGCTCTAAATCGGGGGCTCCC 2580
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2581 TTTAGGGTTCGGATTTAGTGCTTTACGGCACCTCGACCCCAAAAACTTGATTAGGGTGA 2640
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2641 TGGTTCACGTAGTGGGCCATCGCCCTGATAGACGGTTTTTCGCCCTTTGACGTTGGAGTC 2700
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2701 CACGTTCTTTAATAGTGGACTCTTGTTCCAAACTGGAACAACACTCAACCCTATCTCGGT 2760
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 SV40 promoter(2873,3194)
 |
 2821 GATTTAACAAAAATTTAACGCGAATTAATTCTGTGGAATGTGTGTCAGTTAGGGTGTGGA 2880
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2881 AAGTCCCCAGGCTCCCCAGCAGGCAGAAGTATGCAAAGCATGCATCTCAATTAGTCAGCA 2940
 TTCAGGGGTCCGAGGGGTCGTCCGTCTTCATACGTTTCGTACGTAGAGTTAATCAGTCGT

2941 ACCAGGTGTGGAAGTCCCCAGGCTCCCCAGCAGGCAGAAGTATGCAAAGCATGCATCTC 3000
 TGGTCCACACCTTTCAGGGGTCCGAGGGGTCGTCCGTCTTCATACGTTTCGTACGTAGAG

SV40 origin(3040,3117)
|

3001 AATTAGTCAGCAACCATAGTCCCGCCCCTAACTCCGCCCATCCCGCCCCTAACTCCGCC 3060
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3061 AGTTCCGCCCATTTCTCCGCCCATGGCTGACTAATTTTTTTTTATTTATGCAGAGGCCGAG 3120
TCAAGGCGGGTAAGAGGCGGGGTACCGACTGATTAATAAATAAATACGTCTCCGGCTC

3121 GCCGCTCTGCCTCTGAGCTATTCCAGAAGTAGTGAGGAGGCTTTTTTGGAGGCCTAGGC 3180
CGGCGGAGACGGAGACTCGATAAGGTCTTCATCACTCCTCCGAAAAACCTCCGGATCCG

3181 TTTTGCAAAAAGCTCCCGGGAGCTTGTATATCCATTTTCGGATCTGATCAAGAGACAGGA 3240
AAAACGTTTTTCGAGGGCCCTCGAACATATAGGTAAAAGCCTAGACTAGTTCTCTGTCT
Neomycin^R (3256,4050)
|

3241 TGAGGATCGTTTTGCATGATTGAACAAGATGGATTGCACGCAGGTTCTCCGGCCGCTTGG 3300
ACTCCTAGCAAAGCGTACTAACTTGTCTACCTAACGTGCGTCCAAGAGGCCGGCGAACC

3301 GTGGAGAGGCTATTCCGGCTATGACTGGGCACAACAGACAATCGGCTGCTCTGATGCCGCC 3360
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3361 GTGTTCCGGCTGTCAGCGCAGGGGCGCCCGGTTCTTTTTGTCAAGACCGACCTGTCCGGT 3420
CACAAGGCCGACAGTCGCGTCCCCGCGGGCCAAGAAAAACAGTTCTGGCTGGACAGGCCA

3421 GCCCTGAATGAACTGCAGGACGAGGCAGCGCGGCTATCGTGGCTGGCCACGACGGGCGTT 3480
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3481 CCTTGCGCAGCTGTGCTCGACGTTGTCACTGAAGCGGGAAGGGACTGGCTGCTATTGGGC 3540
GGAACGCGTCGACACGAGCTGCAACAGTGACTTCGCCCTTCCCTGACCGACGATAACCCG

3541 GAAGTGCCGGGGCAGGATCTCCTGTCACTCACCTTGCTCCTGCCGAGAAAGTATCCATC 3600
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3601 ATGGCTGATGCAATGCGGCGGCTGCATACGCTTGATCCGGCTACCTGCCCATTCGACCAC 3660
TACCGACTACGTTACGCCGCCGACGTATGCGAACTAGGCCGATGGACGGGTAAGCTGGTG

3661 CAAGCGAAACATCGCATCGAGCGAGCACGTA CTGGATGGAAGCCGGTCTTGTGATCAG 3720
GTTTCGCTTTGTAGCGTAGCTCGCTCGTGCATGAGCCTACCTTCGGCCAGAACAGCTAGTC

3721 GATGATCTGGACGAAGAGCATCAGGGGCTCGCGCCAGCCGAACTGTTCCGACGGCTCAAG 3780
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3781 GCGCGCATGCCCGACGGCGAGGATCTCGTCTGACCCATGGCGATGCCTGCTTGCCGAAT 3840
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3841 ATCATGGTGGAAAATGGCCGCTTTTCTGGATTTCATCGACTGTGGCCGGCTGGGTGTGGCG 3900
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3901 GACCGCTATCAGGACATAGCGTTGGCTACCCGTGATATTGCTGAAGAGCTTGGCGGCGAA 3960
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3961 TGGGCTGACCGCTTCCCTCGTGCTTTACGGTATCGCCGCTCCCGATTTCGAGCGCATCGCC 4020
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4021 TTCTATCGCCTTCTTGACGAGTTCTTCTGAGCGGGACTCTGGGGTTCGAAATGACCGACC 4080
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4081 AAGCGACGCCAACCTGCCATCACGAGATTTTCGATTCCACCGCCGCCTTCTATGAAAGGT 4140
TTCGCTGCGGGTTGGACGGTAGTGCTCTAAAGCTAAGGTGGCGGCGGAAGATACTTTCCA

4141 TGGGCTTCGGAATCGTTTTCCGGGACGCCGGCTGGATGATCCTCCAGCGCGGGGATCTCA 4200
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4201 TGCTGGAGTTCTTCGCCACCCCAACTTGTTTATTGCAGCTTATAATGGTTACAAATAAA 4260
ACGACCTCAAGAAGCGGGTGGGGTTGAACAAATAACGTCGAATATTACCAATGTTTATTT

4261 GCAATAGCATCACAAATTTACAAATAAAGCATTTTTTTTCACTGCATTCTAGTTGTGGTT 4320
CGTTATCGTAGTGTTTAAAGTGTTTATTTTCGTAAAAAAGTGACGTAAGATCAACACCAA

4321 TGTCCAAACTCATCAATGTATCTTATCATGTCTGTATACCGTCGACCTCTAGCTAGAGCT 4380
ACAGGTTTGAGTAGTTACATAGAATAGTACAGACATATGGCAGCTGGAGATCGATCTCGA

4381 TGGCGTAATCATGGTCATAGCTGTTTCCCTGTGTGAAAATTGTTATCCGCTCACAATTCCAC 4440
ACCGCATTAGTACCAGTATCGACAAAGGACACACTTTAACAATAGGCGAGTGTTAAGGTG
lac promoter (4443, 4472)

4441 ACAACATACGAGCCGGAAGCATAAAGTGTAAGCCTGGGGTGCCTAATGAGTGAGCTAAC 4500
TGTTGTATGCTCGGCCTTCGTATTTACATTTTCGGACCCACGGATTACTCACTCGATTG

4501 TCACATTAATTGCGTTGCGCTCACTGCCCGCTTTCAGTCGGGAAACCTGTCGTGCCAGC 4560
AGTGTAATTAACGCAACGCGAGTGACGGGCGAAAGGTCAGCCCTTTGGACAGCACGGTCCG

4561 TGCATTAATGAATCGGCCAACGCGCGGGGAGAGGCGGTTTGCATATTGGGCGCTCTTCCG 4620
ACGTAATTACTTAGCCGGTTGCGCGCCCCTCTCCGCCAACGCATAACCCGCGAGAAGGC

4621 CTTCTCGCTCACTGACTCGCTGCGCTCGGTCGTTTCGGCTGCGGCGAGCGGTATCAGCTC 4680
GAAGGAGCGAGTGACTGAGCGACGCGAGCCAGCAAGCCGACGCCGCTCGCCATAGTCGAG

4681 ACTCAAAGGCGGTAATACGGTTATCCACAGAATCAGGGGATAACGCAGGAAAGAACATGT 4740
TGAGTTTCCGCCATTATGCCAATAGGTGTCTTAGTCCCCTATTGCGTCCTTTCTTGTACA
pBR322 origin (4781, 5397)

4741 GAGCAAAAAGGCCAGCAAAAAGGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCC 4800
CTCGTTTTCCGGTCGTTTTTCCGGTCCCTGGCATTTTTTCCGGCGCAACGACCGCAAAAAGG

4801 ATAGGCTCCGCCCCCTGACGAGCATCACAAAATCGACGCTCAAGTCAGAGGTGGCGAA 4860
TATCCGAGGCGGGGGACTGCTCGTAGTGTTTTTAGCTGCGAGTTCAGTCTCCACCGCTT

4861 ACCCGACAGGACTATAAAGATACCAGGCGTTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTC 4920
TGGGCTGTCTGATATTTCTATGGTCCGCAAAGGGGGACCTTCGAGGGAGCACGCGAGAG

4921 CTGTTCCGACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTGG 4980
GACAAGGCTGGGACGGCGAATGGCCTATGGACAGGCGGAAAGAGGGAAGCCCTTCGCACC

4981 CGCTTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTTCGCTCCAAGC 5040
GCGAAAAGAGTATCGAGTGCGACATCCATAGAGTCAAGCCACATCCAGCAAGCGAGGTTCC

5041 TGGGCTGTGTGCACGAACCCCCCGTTTCAGCCCGACCGCTGCGCCTTATCCGGTAACTATC 5100
 ACCCGACACACGTGCTTGGGGGGCAAGTCGGGCTGGCGACGCGGAATAGGCCATTGATAG

5101 GTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCCACTGGTAACA 5160
 CAGAACTCAGGTTGGGCCATTCTGTGCTGAATAGCGGTGACCGTTCGTCGGTGACCATTTGT

5161 GGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAGTGGTGGCCTAACT 5220
 CCTAATCGTCTCGCTCCATACATCCGCCACGATGTCTCAAGAACTTCACCACCGGATTGA

5221 ACGGCTACACTAGAAGAACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTACCTTCG 5280
 TGCCGATGTGATCTTCTTGTCTATAAACCATAGACGCGAGACGACTTCGGTCAATGGAAGC

5281 GAAAAAGAGTTGGTAGCTCTTGATCCGGCAAACAAACCACCGCTGGTAGCGTTTTTTTTG 5340
 CTTTTTCTCAACCATCGAGAACTAGGCCGTTTGTGGTGGCGACCATCGCCAAAAAAAC

5341 TTTGCAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAAGAAGATCCTTTGATCTTTT 5400
 AAACGTTTCGTCGTCTAATGCGCGTCTTTTTTTTCTAGAGTTCTTCTAGGAACTAGAAAA

5401 CTACGGGGTCTGACGCTCAGTGGAAACGAAAACCTCACGTTAAGGGATTTTGGTCATGAGAT 5460
 GATGCCCCAGACTGCGAGTCACCTTGCTTTTGTAGTGCAATTCCTAAAACAGTACTCTA

5461 TATCAAAAAGGATCTTCACCTAGATCCTTTTAAATTAATAATGAAGTTTAAATCAATCT 5520
 ATAGTTTTTCTTAGAAGTGGATCTAGGAAAATTTAATTTTTACTTCAAATTTAGTTAGA
 Ampillin^R (5552,6412)

5521 AAAGTATATATGAGTAAACTTGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCTA 5580
 TTTTCATATATACTCATTTGAACCAGACTGTCAATGGTTACGAATTAGTCACTCCGTGGAT

5581 TCTCAGCGATCTGTCTATTTTCGTTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAA 5640
 AGAGTCGCTAGACAGATAAAGCAAGTAGGTATCAACGGACTGAGGGGCAGCACATCTATT

5641 CTACGATACGGGAGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCAC 5700
 GATGCTATGCCCTCCCGAATGGTAGACCGGGGTACGACGTTACTATGGCGCTCTGGGTG

5701 GCTCACCGGCTCCAGATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAA 5760
 CGAGTGGCCGAGGTCTAAATAGTCGTTATTTGGTTCGGTCGGCCTTCCCGGCTCGCGTCTT

5761 GTGGTCCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAGCTAGAG 5820
 CACCAGGACGTTGAAATAGGCGGAGGTAGGTACAGATAATTAACAACGGCCCTTCGATCTC

5821 TAAGTAGTTCGCCAGTTAATAGTTTTCGCAACGTTGTTGCCATTGCTACAGGCATCGTGG 5880
 ATTCATCAAGCGGTCAATTATCAAACGCGTTGCAACAACGGTAACGATGTCCGTAGCACC

5881 TGTCACGCTCGTCGTTTGGTATGGCTTCATTCAGCTCCGGTTCCCAACGATCAAGGCGAG 5940
 ACAGTGCAGCAGCAAACCATAACCGAAGTAAGTCGAGGCCAAGGGTTGCTAGTTCCGCTC

5941 TTACATGATCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTCTCCGATCGTTG 6000
 AATGTACTAGGGGGTACAACACGTTTTTTTCGCCAATCGAGGAAGCCAGGAGGCTAGCAAC

6001 TCAGAAAGTAAGTTGGCCGAGTGTTATCACTCATGGTTATGGCAGCACTGCATAATTCTC 6060
 AGTCTTCATTCAACCGGCGTCACAATAGTGAGTACCAATACCGTTCGTGACGTATTAAGAG

6061 TTACTGTCATGCCATCCGTAAGATGCTTTTCTGTGACTGGTGAGTACTCAACCAAGTCAT 6120
 AATGACAGTACGGTAGGCATTCTACGAAAAGACACTGACCACTCATGAGTTGGTTCAGTA

6121 TCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTTGCCCGGCGTCAATACGGGATAATA 6180
 AGACTCTTATCACATACGCCGCTGGCTCAACGAGAACGGGCCGAGTTATGCCCTATTAT

6181 CCGCGCCACATAGCAGAACTTTAAAAGTGCTCATCATTTGGAAAACGTTCTTCGGGGCGAA 6240
 GCGCGGTTGTATCGTCTTGAAATTTTCACGAGTAGTAACCTTTTGCAAGAAGCCCCGCTT

6241 AACTCTCAAGGATCTTACCGCTGTTGAGATCCAGTTCGATGTAACCCACTCGTGCACCCA 6300
 TTGAGAGTTCCTAGAATGGCGACAACCTTAGGTCAAGCTACATTGGGTGAGCACGTGGGT

6301 ACTGATCTTCAGCATCTTTTACTTTTACCAGCGTTTCTGGGTGAGCAAAAACAGGAAGGC 6360
 TGACTAGAAGTCGTAGAAAATGAAAGTGGTCGCAAAGACCCACTCGTTTTTGTCTTCCG

6361 AAAATGCCGCAAAAAAGGGAATAAGGGCGACACGGAAATGTTGAATACTCATACTCTTCC 6420
 TTTTACGGCGTTTTTTCCCTTATTCCCGCTGTGCCTTTACAACCTTATGAGTATGAGAAGG

6421 TTTTTCAATATTATTGAAGCATTATCAGGGTTATTGTCTCATGAGCGGATACATATTTG 6480
 AAAAAGTTATAATAACTTCGTAAATAGTCCCAATAACAGAGTACTCGCCTATGTATAAAC

6481 AATGTATTTAGAAAAATAAACAAATAGGGGTTCGCGCACATTTCCCCGAAAAGTGCCAC 6540
 TTACATAAATCTTTTTATTGTTTATCCCCAAGGCGCGTGTAAAGGGGCTTTTTCACGGTG

6541 CTGACGTC 6548
 GACTGCAG