

***Borrelia burgdorferi*, Signature-Tagged Mutagenesis Library Clone T05TC025 (Gene BB_0629)**

Catalog No. NR-23032

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Contributor:

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Manufacturer:

BEI Resources

Product Description:

Bacteria Classification: *Borreliaceae* (previously *Spirochaetaceae*)¹, *Borrelia*

Species: *Borrelia burgdorferi*

Strain: B31, clone 5A18NP1

Signature-Tagged Mutagenesis Library Clone: T05TC025

Replicon: Chromosome

Gene: BB_0629 [phosphotransferase system (PTS), fructose-specific IIABC component]

Insertion Site^{2,3}: 611156

Original Source: *Borrelia burgdorferi* (*B. burgdorferi*), clone T05TC025 was produced by signature-tagged mutagenesis (STM) of the BB_0629 gene.^{2,3}

Comments: *B. burgdorferi*, strain B31 5A18NP1 STM library clone T05TC025 lacks linear plasmids lp28-4 and lp56. The plasmid profile was determined by PCR using plasmid specific primers.³

B. burgdorferi is a Gram-negative, motile spirochete.⁴ It is a zoonotic, vector-borne pathogen transmitted by ticks and the etiologic agent of Lyme disease, now the most common tick-transmitted disease in the United States.⁵ *B. burgdorferi* is predominant in North America, but also exists in Europe.

B. burgdorferi, strain B31 was originally isolated in 1981 from adult ticks (*Ixodes dammini*) collected from lower vegetation on Shelter Island, New York, USA.^{4,5} Strain B31 is composed of a 910 kilobase (kb) linear chromosome, 9 circular plasmids (cp) and 12 linear plasmids (lp). Plasmids range in size from 5 kb to 56 kb and total 610 kb.^{3,6} Continuous passage of *B. burgdorferi* is known to cause spontaneous loss of plasmids. The complete genome of *B. burgdorferi*, strain B31 has been sequenced (GenBank: [AE000783](https://www.ncbi.nlm.nih.gov/nuccore/AE000783)).⁷

B. burgdorferi, strain B31, clone 5A18NP1 was derived from the low-passage clone 5A18 of strain B31.⁸ Clone 5A18NP1 lacks lp56 and lp28-4 and the BBE02 gene (a putative restriction-modification gene on lp25) was disrupted by homologous recombination resulting in kanamycin resistance.⁹ Inactivation of BBE02 results in increased transformation efficiency and therefore, clone 5A18NP1, was

used to create the STM library through the *mariner*-based transposition suicide *Himar1* delivery vector, pMarGent, containing *aacC1* which confers gentamicin resistance.^{2,3,10} STM is a negative selection method that identifies clones by unique DNA sequences that are incorporated into the transposable element.³

Material Provided:

Each vial contains approximately 0.5 mL of bacterial culture in Revised Barbour-Stoenner-Kelly medium supplemented with 200 µg/mL kanamycin, 40 µg/mL gentamicin and 15% glycerol.

Note: If homogeneity is required for your intended use, please purify prior to initiating work.

Packaging/Storage:

NR-23032 was packaged aseptically, in screw-capped plastic cryovials. The product is provided frozen and should be stored at -80°C or colder immediately upon arrival. For long-term storage, the vapor phase of a liquid nitrogen freezer is recommended. Freeze-thaw cycles should be avoided.

Growth Conditions:

Media:

Revised Barbour-Stoenner-Kelly broth (see Appendix I) with 200 µg/mL kanamycin and 40 µg/mL gentamicin

Revised Barbour-Stoenner-Kelly agar (see Appendix I) with 200 µg/mL kanamycin, 40 µg/mL gentamicin and 0.8% agar

Incubation:

Temperature: 32°C to 34°C (growth at 37°C may result in plasmid loss)²

Atmosphere: Microaerophilic (slower growth occurs under aerobic conditions²)

Propagation:

1. Keep vial in dry ice during inoculations.
2. Inoculate new cultures from scraping of frozen stock into a single tube of Revised Barbour-Stoenner-Kelly Medium.
3. Incubate the tube at 32 to 34°C for 2 to 14 days. Do not shake culture during growth.

Note: Subculturing should be minimized to avoid plasmid loss.^{2,8}

Citation:

Acknowledgment for publications should read "The following reagent was obtained through BEI Resources, NIAID, NIH: *Borrelia burgdorferi*, Signature-Tagged Mutagenesis Library Clone T05TC025 (Gene BB_0629), NR-23032."

Biosafety Level: 2

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the following publication: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, and National Institutes of Health. Biosafety in Microbiological and Biomedical Laboratories, 5th ed. Washington, DC: U.S. Government Printing Office, 2009; see www.cdc.gov/biosafety/publications/bmbl5/index.htm.

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References:

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for Characterization of the Phylum *Spirochaetes* and Its Major Clades: Proposal for a Taxonomic Revision of the Phylum." *Front. Microbiol.* 4 (2013): 217. Erratum in: *Front. Microbiol.* 4 (2013): 322. PubMed: 23908650.

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3. Lin, T., et al. "Analysis of an Ordered, Comprehensive STM Mutant Library in Infectious *Borrelia burgdorferi*: Insights into the Genes Required for Mouse Infectivity." *PLoS One* 7 (2012): e47532. PubMed: 23133514.

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5. Johnson, R. C., et al. "*Borrelia burgdorferi* sp. nov.: Etiologic Agent of Lyme Disease." *Int. J. Syst. Bacteriol.* 34 (1984): 496-497.

6. Casjens, S., et al. "A Bacterial Genome in Flux: The Twelve Linear and Nine Circular Extrachromosomal DNAs in an Infectious Isolate of the Lyme Disease Spirochete *Borrelia burgdorferi*." *Mol. Microbiol.* 35 (2000): 490-516. PubMed: 10672174.

7. Fraser, C. M., et al. "Genomic Sequence of a Lyme Disease Spirochaete, *Borrelia burgdorferi*." *Nature* 390 (1997): 580-586. PubMed: 9403685.

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Appendix I: Revised BSK Medium (ATCC® Medium: 1914)

HEPES.....	5.64 g
Neopeptone.....	4.7 g
Sodium citrate.....	0.7 g
Glucose.....	5.64 g
NaHCO ₃	2.0 g
TC-Yeastolate	2.0 g
Sodium pyruvate.....	0.75 g
N-acetylglucosamine.....	0.37 g
Bovine serum albumin, fraction V.....	47.0 g
CMRL 1066, 10X (w/o Glutamine or NaHCO ₃).....	100.0 mL
Rabbit serum (heat inactivated).....	60.0 mL
Distilled water.....	840 mL

For agar, add 0.8% agarose.

Dissolve ingredients up to and including bovine serum albumin one at a time in distilled water. Adjust to pH 7.5 with NaOH and filter-sterilize. Aseptically add CMRL 1066 and rabbit serum. Mix well and aseptically dispense into appropriate vessel. Final pH of complete medium should be 7.5 - 7.6.