

***Plasmodium berghei*, Strain NK65 RedStar**

Catalog No. MRA-905

For research use only. Not for human use.

Contributor:

Ute Frevert, D.V.M., Ph.D., Associate Professor, Department of Microbiology (Parasitology), New York University School of Medicine, New York, New York, USA

Manufacturer:

BEI Resources

Product Description:

Protozoa Classification: *Plasmodiidae*, *Plasmodium*

Species: *Plasmodium berghei*

Strain: NK65 RedStar

Original Source: *Plasmodium berghei* (*P. berghei*), strain NK65 RedStar is a clone of *P. berghei*, strain NK65.^{1,2} *P. berghei*, strain NK65 was isolated in April 1964 from *Anopheles durenii millecampi* mosquitoes collected in the River Kisanga, Democratic Republic of Congo.³

Comment: *P. berghei*, strain NK65 RedStar was deposited as a pyrimethamine-resistant parasite that stably expresses a red fluorescent protein (RFP), RedStar, during sporozoite stages.^{1,2}

P. berghei is a protozoan parasite that infects mammals other than humans, especially rodents. It is commonly used in rodent model studies of malaria.⁴

Material Provided:

Each vial of MRA-905 contains approximately 0.5 mL of *P. berghei*, strain NK65 RedStar infected mouse blood in Glycerolyte 57 solution (1:2). Please see Appendix I for cryopreservation instructions.

Packaging/Storage:

MRA-905 was packaged aseptically in cryovials. The product is provided frozen and should be stored at -80°C or colder immediately upon arrival. For long-term storage, the vapor phase of a liquid nitrogen freezer is recommended (-130°C or colder). Freeze-thaw cycles should be avoided.

Growth Conditions⁵:

Thaw a frozen cryovial of MRA-905 in a 35°C to 37°C water bath for approximately 2 to 3 minutes. Do not allow the vial to immerse near the cap line seal while thawing. Once thawed, wipe the outside of the vial with 70% to 100% ethanol before opening. Using a 27 gauge 1/2 inch needle, inject thawed parasites into mice via intraperitoneal injection (IP) at 50 µL to 100 µL per mouse. Monitor growth of parasites by tail vein bleed sampling and Giemsa-stained thin blood smear microscopy daily, starting on day 3 post-inoculation. Harvest and/or serially passage infected blood

to naive mice when parasitemia reaches 1% to 5%, before significant signs of infection appear. To maintain the parasite strain *in vivo*, passage infected blood from a donor mouse to a recipient mouse using intraperitoneal or intravenous (IV) injection of an appropriately diluted inoculum as required for the study.

Note: Do not directly inject freshly thawed parasites from cryopreserved stocks by the IV route, as these samples contain cryoprotectant, anticoagulant and may contain traces of lysed or coagulated red blood cells. Direct IV inoculation from cryopreserved stock may result in pulmonary embolism or shock in mice.

Citation:

Acknowledgment for publications should read “The following reagent was obtained through BEI Resources, NIAID, NIH: *Plasmodium berghei*, Strain NK65 RedStar, MRA-905, contributed by Ute Frevert.”

Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the following publication: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, and National Institutes of Health. Biosafety in Microbiological and Biomedical Laboratories. 5th ed. Washington, DC: U.S. Government Printing Office, 2009; see www.cdc.gov/biosafety/publications/bmbl5/index.htm.

All blood cultures should be handled with appropriate safety precautions necessary for the handling of bloodborne pathogens. Personnel must be trained in accordance with their institutional policy regarding bloodborne pathogens.

Disclaimers:

You are authorized to use this product for research use only. It is not intended for human use.

Use of this product is subject to the terms and conditions of the BEI Resources Material Transfer Agreement (MTA). The MTA is available on our Web site at www.beiresources.org.

While BEI Resources uses reasonable efforts to include accurate and up-to-date information on this product sheet, neither ATCC® nor the U.S. Government makes any warranties or representations as to its accuracy. Citations from scientific literature and patents are provided for informational purposes only. Neither ATCC® nor the U.S. Government warrants that such information has been confirmed to be accurate.

This product is sent with the condition that you are responsible for its safe storage, handling, use and disposal. ATCC® and the U.S. Government are not liable for any damages or injuries arising from receipt and/or use of this product. While reasonable effort is made to ensure authenticity and reliability of materials on deposit, the U.S.

Government, ATCC®, their suppliers and contributors to BEI Resources are not liable for damages arising from the misidentification or misrepresentation of products.

Use Restrictions:

This material is distributed for internal research, non-commercial purposes only. This material, its product or its derivatives may not be distributed to third parties. Except as performed under a U.S. Government contract, individuals contemplating commercial use of the material, its products or its derivatives must contact the contributor to determine if a license is required. U.S. Government contractors may need a license before first commercial sale.

References:

1. Frevert, U., Personal Communication.
2. Frevert, U., et al. "Intravital Observation of *Plasmodium berghei* Sporozoite Infection of the Liver." PLoS Biol. 3 (2005): e192. PubMed: 15901208.
3. Ramiro, R. S., S. E. Reece and D. J. Obbard. "Molecular Evolution and Phylogenetics of Rodent Malaria Parasites." BMC Evol. Biol. 12 (2012): 219. PubMed: 23151308.

4. Hall, N., et al. "A Comprehensive Survey of the *Plasmodium* Life Cycle by Genomic, Transcriptomic, and Proteomic Analyses." Science 307 (2005): 82-86. PubMed: 15637271.
5. Peters, W. and B. L. Robinson (1999), "Chapter 92 -- Malaria." In Handbook of Animal Models of Infection. Eds. O. Zak and M. Sande, Academic Press: London, pp. 757-773.
6. Engelmann, S., P. Sinnis and K. Matuschewski. "Transgenic *Plasmodium berghei* Sporozoites Expressing Beta-Galactosidase for Quantification of Sporozoite Transmission." Mol. Biochem. Parasitol. 146 (2006): 30-37. PubMed: 16316690.
7. Knop, M., et al. "Improved Version of the Red Fluorescent Protein (drFP583/DsRed/RFP)." Biotechniques 33 (2002): 592-602. PubMed: 12238769.

ATCC® is a trademark of the American Type Culture Collection.



APPENDIX I: CRYOPRESERVATION

Note: Only immature blood-stage parasites (rings) are viable by this method. Parasitemia of 1% or higher of ring-stage parasites is recommended for cryopreservation. All steps should be carried out in a biosafety cabinet under proper air flow.

1. Harvest parasitized mouse blood into 25 x volume ice cold sterile PBS (pH ~ 7.2) containing an appropriate concentration of anticoagulant (e.g. Heparin at 10 units/mL) and place on ice.
2. Centrifuge the diluted blood culture at 1000 x g for 5 minutes at 4°C.
3. Aspirate the supernatant carefully. Measure the volume of packed red blood cells using centrifuge tube graduations or standard volume controls.
4. To the volume of packed red blood cells, add dropwise one volume of cold (4°C) Glycerolyte 57 solution. Let stand for 5 minutes at room temperature.
5. Add dropwise an additional volume of cold Glycerolyte 57 solution to the pellet. Mix well and aliquot 0.5 mL into 1.5 mL sterile cryopreservation vials.
6. Place the vials in a controlled-rate freezing unit. From room temperature, cool the vials at -1°C/min to -40°C. If the freezing unit can compensate for the heat of fusion, maintain rate at -1°C/min through this phase. At -40°C, plunge vials into liquid nitrogen. Alternatively, place the vials in a Nalgene 1°C freezing container. Place the container at -80°C for 24 to 48 hours and then plunge vials into liquid nitrogen.
7. Store in either the vapor or liquid phase of a nitrogen refrigerator (-130°C or colder).