

## Plasmodium falciparum, Strain HOX

Catalog No. MRA-1275

For research use only. Not for human use.

### Contributor:

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### Manufacturer:

BEI Resources

### Product Description:

Protozoa Classification: Plasmodiidae, Plasmodium

Species: Plasmodium falciparum

Strain: HOX (High OXYgen)

Original Source: Plasmodium falciparum (P. falciparum), strain HOX is a derivative of the NF54 strain that was gradually adapted to proliferate in high oxygen conditions (standard tissue culture) of 5% CO<sub>2</sub> in air at 37°C.<sup>1</sup> The parent P. falciparum, strain NF54 (available as BEI Resources MRA-1000) was isolated from a patient living near Schipol Airport, Amsterdam, who had never left the Netherlands.<sup>2</sup>

Comments: P. falciparum, strain HOX is reported to be sensitive to artemisinin, atovaquone, chloroquine, doxycycline and pyrimethamine.<sup>1</sup> The whole genome shotgun sequence of P. falciparum, strain NF54 is available (GenBank: [AMYQ000000000](#)).

### Material Provided:

Each vial of MRA-1275 contains approximately 0.5 mL of P. falciparum infected human blood in Glycerolyte 57 solution (1:5). Please see Appendix I for cryopreservation instructions.

### Packaging/Storage:

MRA-1275 was packaged aseptically in cryovials. The product is provided frozen and should be stored at -80°C or colder immediately upon arrival. For long-term storage, the vapor phase of a liquid nitrogen freezer is recommended (-130°C or colder). Freeze-thaw cycles should be avoided.

### Growth Conditions:

RPML 1640 medium (without phenol red and with glutamine), adjusted to contain 25 mM HEPES, 27 mM NaHCO<sub>3</sub>, 4.4 μM hypoxanthine and 0.5% Albumax

Please see Appendix II for complete medium preparation instructions and notes.

### Incubation:

Temperature: 37°C

Atmosphere: Aerobic with 5% CO<sub>2</sub>

### Propagation:

1. Place the frozen vial in a 37°C water bath until the culture is completely thawed. Transfer the vial to a

biological safety hood and wipe the outside surface of the vial with 70% ethanol.

2. Using a sterile 1 mL pipette, aseptically transfer the contents of the vial to a sterile 50 mL conical centrifuge tube.
3. Add 12% sodium chloride (NaCl) solution dropwise, approximately 1:5 ratio NaCl to cell mixture (0.2x original culture volume). Allow to stand for 5 minutes.
4. Using a 1 mL syringe and 27-gauge needle, add dropwise while shaking 10 volumes of a 1.6% NaCl solution (10:1 ratio NaCl to original culture volume).
5. Centrifuge at 1000 × g for 5 minutes and remove most of the supernatant, leaving approximately 0.5 mL to 1 mL to resuspend the cell pellet. Resuspend the cells by gently swirling the tube.
6. Add dropwise while shaking 10 volumes of complete medium. Centrifuge at 1000 × g for 5 minutes and carefully remove the supernatant.
7. Add 10 mL of complete medium and transfer the sample to a 25 cm<sup>2</sup> tissue culture flask with vented cap.
8. For continuous culture, add uninfected red blood cells (RBCs) to a 1% to 2% hematocrit solution (immediately or the next day).
9. Incubate the tissue culture flask at 37°C in 5% CO<sub>2</sub> incubator.
10. Take a smear for Giemsa staining after 24 hours to evaluate parasite growth and determine parasitemia.

### Maintenance:

Note: Changing of the culture medium every 24 hours is required for malaria-infected erythrocyte cultures.

1. Remove the flask with infected culture from the 37°C incubator and place onto a flask warmer.
2. Carefully remove the supernatant with a sterile, unplugged Pasteur pipet under vacuum. Remove as much of the supernatant as possible without taking the cells.
3. Add 25 mL of sterile warm (37°C) complete medium to the flask, then tighten the vented cap and place the flask in the 37°C incubator until the next change of medium.

### Preparation of Blood Smear:

1. Carefully remove 0.5 mL to 1 mL of mixed culture with a sterile pipet and transfer to a microcentrifuge tube.
2. Centrifuge the microcentrifuge tube at high speed and aspirate the supernatant.
3. Mix the pellet and transfer 6 μL of the suspension to a glass slide for a thick film smear or 2 μL for a thin film smear. Spread the drop into a thin film using the edge of a clean glass slide. Air dry for 3 minutes at room temperature.
4. Fix the blood smear by rinsing it with methyl alcohol. Air dry for 3 minutes at room temperature.
5. Stain blood films in 5% Giemsa solution for 15 minutes. Rinse with distilled water and allow to air dry.
6. Using light microscopy at 100x magnification, determine parasitemia of culture.

### Citation:

Acknowledgment for publications should read "The following reagent was obtained through BEI Resources, NIAID, NIH:

# Product Information Sheet for MRA-1275

*Plasmodium falciparum*, Strain HOX, MRA-1275, contributed by Theresa A. Shapiro."

## Biosafety Level: 2

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the following publication: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, and National Institutes of Health. Biosafety in Microbiological and Biomedical Laboratories. 5th ed. Washington, DC: U.S. Government Printing Office, 2009; see [www.cdc.gov/biosafety/publications/bmbl5/index.htm](http://www.cdc.gov/biosafety/publications/bmbl5/index.htm).

All blood cultures should be handled with appropriate safety precautions necessary for the handling of bloodborne pathogens. Personnel must be trained in accordance with their institutional policy regarding bloodborne pathogens.

## Disclaimers:

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## References:

1. Bakshi, R. P., et al. "Model System to Define Pharmacokinetic Requirements for Antimalarial Drug

Efficacy." Sci. Transl. Med. 5 (2013): 205ra135. PubMed: 24089407.

2. Ponnudurai, T., A. D. Leeuwenberg and J. H. Meuwissen. "Chloroquine Sensitivity of Isolates of *Plasmodium falciparum* Adapted to *In Vitro* Culture." Trop. Geogr. Med. 33 (1981): 50-54. PubMed: 7018038.

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## APPENDIX I: CRYOPRESERVATION

**Note:** Only the immature parasite stage (rings) is viable by this method. We recommend a parasitemia of 3% or higher of ring-stage parasites for cryopreservation.

1. Centrifuge the culture at 1000 × g for 5 minutes.
2. Wash the pellet once with 10 or more volumes of incomplete RPMI 1640 medium. Centrifuge at 1800 × g for 5 minutes and leave sufficient supernatant to resuspend the pellet.
3. To the volume of packed red blood cells, slowly add dropwise one volume of cold (4°C) Glycerolyte 57 solution. Let stand for 5 minutes at room temperature.
4. Add dropwise an additional 3 volumes of cold Glycerolyte 57 solution to the pellet. Mix well and aliquot 0.5 mL into 1.5 mL sterile cryopreservation vials.
5. Place the vials in a controlled-rate freezing unit. From room temperature, cool the vials at -1°C/min to -40°C. If the freezing unit can compensate for the heat of fusion, maintain rate at -1°C/min through this phase. At -40°C, plunge vials into liquid nitrogen. Alternatively, place the vials in a Nalgene 1°C freezing container. Place the container at -80°C for 24 to 48 hours and then plunge vials into liquid nitrogen.
6. Store in either the vapor or liquid phase of a nitrogen refrigerator (-130°C or colder).

## APPENDIX II: MEDIA PREPARATION

1. **Incomplete Medium:** Used for many applications involving wash steps during preparation of parasites for culture or assay. The incomplete medium consists of RPMI 1640 medium supplemented with the following components<sup>1</sup>:

### Incomplete Medium

RPMI 1640 medium<sup>2,3</sup>

Sodium bicarbonate (NaHCO<sub>3</sub>)<sup>4</sup> 2.0 g/L

L-Glutamine 2 mM

HEPES [4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid] 25 mM

### Optional:

D-Glucose<sup>5</sup> 2.0 g/L

Hypoxanthine 5 µg/L

Gentamycin 2.5 mg/L

<sup>1</sup>Prepare sterile stock solutions at concentrations that are easily diluted into the liquid medium to obtain the appropriate user concentrations, and add aseptically. Ready-made stock solutions for many of the components are available from numerous manufacturers.

<sup>2</sup>RPMI 1640 medium is available from numerous manufacturers as both a powder and a sterile, prepared liquid, with or without L-glutamine and HEPES. If using powdered RPMI 1640 medium, prepare the medium following manufacturer instructions, sterile-filter using a 0.22 µm filter, then aseptically add the necessary components in the appropriate concentrations.

<sup>3</sup>If stock solutions were not sterile or aseptic techniques were not followed, sterile-filter the medium using a 0.22 µm filter after the addition of all components. Store at 4°C.

<sup>4</sup>Prepared, liquid medium typically contains sodium bicarbonate while powdered medium does not. A typical concentration of sodium bicarbonate in RPMI 1640 medium is 2 g/L, though some formulations contain different amounts.

<sup>5</sup>A typical concentration of D-glucose in RPMI 1640 medium is 2 g/L. The option to supplement with an additional 2 g/L yields a final concentration of 4 g/L D-glucose.

2. **Complete Medium:** Consists of incomplete medium (above) supplemented with 0.5% Albumax. If necessary, filter the complete medium with a 0.22 µm filter. Since serum tends to clog sterilizing filters, a serum pre-filter may be used first, followed by a 0.22 µm sterilizing filter.

**Note:** Human serum is not required. Albumax is used with washed type O blood.