

**Helicobacter pylori, Strain Hp H-11**

**Catalog No. NR-43683**

**Product Description:** *Helicobacter pylori* (*H. pylori*), strain Hp H-11 was isolated from gastric biopsy homogenate of a patient with gastritis in Ohio, USA.

**Lot<sup>1</sup>: 64136572**

**Manufacturing Date: 07APR2016**

TEST	SPECIFICATIONS	RESULTS
<b>Phenotypic Analysis</b> Cellular morphology Colony morphology <sup>2</sup>  Motility (wet mount) Biochemical characterization Catalase Oxidase Urease Nitrate reduction H <sub>2</sub> S (lead acetate paper) Hippurate hydrolysis Growth with 5% CO <sub>2</sub> Growth at 25°C Growth at 37°C Growth at 42°C Brucella albimi + 0.16% agar (growth control) Brucella albimi + 0.16% agar with 1% glycine Brucella albimi + 0.16% agar with 3.5% NaCl	Gram-negative rods Report results  Report results  Positive Positive Positive Negative Report results Negative Growth No growth Growth Report results Growth No growth No growth	Gram-negative rods Circular, low convex, entire, translucent and gray (Figure 1) Motile  Positive Positive Positive Negative Positive Negative Growth No growth Growth Growth Growth Growth No growth <sup>3</sup> No growth <sup>4</sup>
<b>Antibiotic Susceptibility Profile</b> BD BBL™ Sensi-Disc™ susceptibility test discs Metronidazole (80 µg) <sup>5</sup> Nalidixic acid (30 µg) <sup>6</sup>	Report results Report results	40 mm 10 mm
<b>Genotypic Analysis</b> Sequencing of 16S ribosomal RNA gene (~ 1390 base pairs)  Digital DNA-DNA hybridization (dDDH) <sup>7</sup>	≥ 99% sequence identity to <i>H. pylori</i> , strain Hp H-11 (GenBank: AKPC01000002.1) > 70% agreement for species identification	100% sequence identity to <i>H. pylori</i> , strain Hp H-11 (GenBank: AKPC01000002.1) <i>H. pylori</i> (92.2%)
<b>Confirmation of <i>H. pylori</i> by PCR Amplification of Extracted DNA</b> Positive control (16S ribosomal RNA gene) Negative control ( <i>H. acinonychis</i> ) <i>ureA</i> <i>ssaA</i>	Amplicon present No amplicon present Amplicon present Amplicon present	Amplicon present No amplicon present No amplicon present <sup>8</sup> Amplicon present
<b>Purity (post-freeze)</b> Microaerophilic growth <sup>9</sup>  Aerobic growth <sup>10,11</sup>	Growth consistent with expected colony morphology Growth consistent with expected colony morphology	Growth consistent with expected colony morphology Growth consistent with expected colony morphology
<b>Viability (post-freeze)<sup>2</sup></b>	Growth	Growth

<sup>1</sup>NR-43683 was produced by inoculation of the deposited material into Brucella broth. Broth inoculum was added to Columbia agar with 7% defibrinated horse blood, 5 µg/mL trimethoprim, 5 µg/mL vancomycin, 10 µg/mL cefsulodin and 2.5 µg/mL amphotericin B. The inoculated agar and broth were each grown for 4 days at 37°C in a microaerophilic atmosphere (~ 6-16% O<sub>2</sub> and 2-10% CO<sub>2</sub>). Colonies from the Columbia agar

- culture were suspended into the Brucella broth growth, and this biphasic culture was added to Columbia agar with 7% defibrinated horse blood, 5 µg/mL trimethoprim, 5 µg/mL vancomycin, 10 µg/mL cefsulodin and 2.5 µg/mL amphotericin B kolles, which were grown for 2 days 37°C in a microaerophilic atmosphere to produce this lot.
- <sup>2</sup>7 days on Columbia agar with 7% defibrinated horse blood, 5 µg/mL trimethoprim, 5 µg/mL vancomycin, 10 µg/mL cefsulodin and 2.5 µg/mL amphotericin B at 37°C in a microaerophilic atmosphere
- <sup>3</sup>Specifications for these tests were obtained from Bergey's Manual® of Systematic Bacteriology, 2<sup>nd</sup> ed., Volume 2, Part C, which indicates that growth may occur in up to 17% of strains.
- <sup>4</sup>Specifications for these tests were obtained from Bergey's Manual® of Systematic Bacteriology, 2<sup>nd</sup> ed., Volume 2, Part C, which indicates that growth may occur in 20% to 43% of strains.
- <sup>5</sup>Test performed using metronidazole 80 µg (MET-80, BBL™ catalog no. 231605)
- <sup>6</sup>Test performed using nalidixic acid 30 µg (NA-30, BBL™ catalog no. 231311)
- <sup>7</sup>Relatedness between bacterial strains has traditionally been determined using DDH. For additional information refer to Auch, A.F., et al. "Digital DNA-DNA Hybridization for Microbial Species Delineation by Means of Genome-to-Genome Sequence Comparison." *Stand Genomic Sci*, 2 (2010): 117-134, PubMed: 21304684.
- <sup>8</sup>PCR amplification of DNA from NR-43683 did not produce the ~380 base pair amplicon corresponding to the *H. pylori*-specific *ureA* gene. Sequence analysis of the *ureA* gene from *H. pylori*, Strain Hp H-11 (GenBank: AKPC00000000.1) identified sequence differences in the binding site for the reverse primer, which negatively affected extension from this primer and resulted in no amplicon being generated.
- <sup>9</sup>Purity of this lot was assessed for 7 days on Tryptic Soy agar with 5% defibrinated sheep blood at 37°C in a microaerophilic atmosphere (~ 6-16% O<sub>2</sub> and 2-10% CO<sub>2</sub>).
- <sup>10</sup>Purity of this lot was assessed for 7 days on Tryptic Soy agar with 5% defibrinated sheep blood at 37°C in an aerobic atmosphere with 5% CO<sub>2</sub>.
- <sup>11</sup>*H. pylori* is known to show weak growth under aerobic conditions (Bury-Moné, S., et al. "Is *Helicobacter pylori* a True Microaerophile?" *Helicobacter* 11 (2006): 296-303. PubMed: 16882333.).

Figure 1: Colony Morphology



Date: 26 OCT 2016

Signature:

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