



NIH AIDS Reagent Program

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DATA SHEET

Reagent: ☒ HIV-1 ASM 3 Virus

Catalog Number: 2792

Lot Number: 14 Apr 95 (D7)

Release Category: A

Provided: Each isolate is provided as 1 ml cell-free virus.

Original Source: These virus stocks originate from clinically and virologically well defined HIV seropositive patients enrolled in the San Francisco AIDS Clinic Cohort. Virus was isolated from either healthy long term positive (HLP) individuals seropositive for >10 years with CD4 counts generally >500/mm³, or from recent seroconverters (RSC) seropositive for 500/mm³.

Host Strain: Human PBMCs

Description: R5.

Special Characteristics: All isolates have been propagated only in PBMCs, and only for a few passages. The viruses may be useful for phenotypic and genotypic characterization of virologic factors that influence the rate of progression of HIV disease.

Recommended Storage: Liquid nitrogen.

Contributor: Drs. Mark Feinberg and Susan Buchbinder.

References: Buchbinder SP, Katz MH, Hessol NA, O'Malley PM, Homberg SD. Long-term HIV-1 infection without immunologic progression. *AIDS* **8**:1123-1128, 1994.

ALL RECIPIENTS OF THIS MATERIAL MUST COMPLY WITH ALL APPLICABLE BIOLOGICAL, CHEMICAL, AND/OR RADIOCHEMICAL SAFETY STANDARDS INCLUDING SPECIAL PRACTICES, EQUIPMENT, FACILITIES, AND REGULATIONS. NOT FOR USE IN HUMANS.

NOTE:

Acknowledgment for publications should read "The following reagent was obtained through the NIH AIDS Reagent Program, Division of AIDS, NIAID, NIH: HIV-1 ASM 3 from Drs. Mark Feinberg and Susan Buchbinder." Also include the reference cited above in any publications.

Virus	Cat. No.	Lot	TCID50/0.2 ml	Type	Co-receptor Usage	Plasma Viral Load (copies/ml)	CD4/mm ³	CD8/mm ³
ASM 79	2784	2 14 Apr 95 (D7)	1:6208	RSC	R5	4460	865	1270
ASM 34	2785	2 5 Apr 95 (D5)	1:57,052	HLP	R5	43,000	375	1922
ASM 93765	2786	2 6 Apr 95 (D6)	1:43,238	HLP	R5	1150	769	915
ASJM 108	2787	2 6 Apr 95 (D6)	1:18,820	HLP	R5	148,000	449	758
ASM 80	2788	2 6 Apr 95 (D6)	1:57,052	RSC	R5	94,000	427	1325
ASM 94122	2789	2 7 Apr 95 (D7)	1:8192	HLP	R5	59,000	553	656
ASM 57	2790	2 13 Apr 95 (D6)	1:32,768	HLP	R5	110,000	1083	1054
ASM 44	2791	2 6 Apr 95 (D6)	1:10,809	RSC	R5X4	203,000	531	1054
ASM 3	2792	2 14 Apr 95 (D7)	1:1552	HLP	R5		1066	956
ASM 3	2792	3 17 Apr 95 (D10)	1:1552	HLP	R5		1066	956
ASM 61	2793	2 6 Apr 95 (D6)	1:24,834	HLP	R5	42,000	886	675
ASM 71	2794	2 14 Apr 95 (D7)	1:2702	HLP	R5	273	934	837
ASM 54	2795	2 13 Apr 95 (D6)	1:4705	RSC	R5X4		483	1154
ASM 121	2796	2 13 Apr 95 (D6)	1:1176	HLP	R5	1193	782	1865
ASM 42	2797	2 13 Apr 95 (D6)	1:10,809	RSC	R5	1227	661	1338
ASM 93534	2798	2 14 Apr 95 (D7)	1:2702	HLP	R5		1378	1378

Propagation of ASM Viruses

Reagent

PHA Medium

RPMI 1640 supplemented with L-glutamine, 20% heat-inactivated fetal bovine serum, 3% natural IL-2 (Boehringer Mannheim or equivalent), 5 µg/ml PHA-P (Sigma or equivalent), and 50 µg/ml gentamicin

Culture Medium

RPMI 1640 supplemented with L-glutamine, 20% heat-inactivated fetal bovine serum, 5% natural IL-2 (Boehringer Mannheim or equivalent), and 50 µg/ml gentamicin

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Propagation

1. Stimulate a mixture of PMBCs from two donors (2×10^6 cells/ml) in PHA Medium for 24 hours.
2. Resuspend donor cells (2×10^6 cells/ml) in Culture Medium. Add 0.5 ml of stock virus to 10 ml of culture medium containing 2×10^7 freshly stimulated PBMCs in a 15 ml culture tube, and incubate overnight.
3. Wash the cells twice with PBS-CMF. Resuspend the cell pellet in 40 ml Culture Medium containing 6×10^7 freshly stimulated PBMCs, and transfer the infected cells to a tissue culture flask.
4. Replace half of the medium with fresh Culture Medium every third day, and with fresh Culture Medium containing freshly stimulated PBMCs every seventh day. Monitor p24 antigen levels every 3-4 days.

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Reagent

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<i>Culture Medium</i>	RPMI 1640 supplemented with L-glutamine, 20% heat-inactivated fetal bovine serum, 5% natural IL-2 (Boehringer Mannheim or equivalent), and 50 µg/ml gentamicin

Propagation

1. Stimulate a mixture of PMBCs from two donors (2×10^6 cells/ml) in PHA Medium for 24 hours.
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Last Updated: July 30, 2018

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