

## Plasmodium berghei, Strain (ANKA) 507m6cl1

Catalog No. MRA-867

For research use only. Not for human use.

### Contributor:

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### Manufacturer:

BEI Resources

### Product Description:

Protozoa Classification: Plasmodiidae, Plasmodium

Species: Plasmodium berghei

Strain: (ANKA) 507m6cl1

Original Source: Plasmodium berghei (P. berghei) strain (ANKA) 507m6cl1 is a genetically modified clone of the ANKA strain following stable transfection with the pL0023 vector (BEI Resources MRA-792). This line expresses the green fluorescent protein (GFP) and has been selected by flow (FACS) sorting, based on GFP fluorescence. This line does not contain a drug-selectable marker.<sup>1,2</sup>

Comments: The parent strain P. berghei, ANKA was isolated in July 1965 from Anopheles durenii millescampi mosquitoes collected in the River Kasapa, Democratic Republic of Congo.<sup>3</sup> The complete genome of P. berghei ANKA, has been sequenced (BioProject: [PRJNA317456](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA317456)).<sup>4,5</sup>

P. berghei is a protozoan parasite that infects mammals other than humans, especially rodents. It is commonly used in rodent model studies of malaria.<sup>3</sup> P. berghei preferentially invades reticulocytes, typically producing infections in mice that induce severe pathology.<sup>4</sup>

The transgene from pL2003 is integrated into the genome by double cross-over integration and therefore parasites should not lose the transgene and will remain fluorescent throughout the life cycle.<sup>1,2</sup> Gametocyte, ookinete, oocysts, sporozoite and liver development is comparable to wildtype P. berghei (ANKA).<sup>2</sup>

### Material Provided:

Each vial contains approximately 0.5 mL of P. berghei-infected mouse blood in Glycerolyte 57 solution (1:2). This item is host restricted and must be amplified in rodents. Please see Appendix I for cryopreservation instructions.

### Packaging/Storage:

MRA-867 was packaged aseptically in cryovials. The product is provided frozen and should be stored at -80°C or colder immediately upon arrival. For long-term storage, the vapor phase of a liquid nitrogen freezer is recommended (-130°C or colder). Freeze-thaw cycles should be avoided.

### Growth Conditions:<sup>6</sup>

*In vivo*, Swiss Webster mouse

Note: Some strains of mice may require dietary or drug pretreatment protocols for successful infection as P. berghei strains have a strong predilection for invasion of reticulocytes.

### Inoculation:

1. Thaw a frozen cryovial of MRA-867 in a 35°C to 37°C water bath for approximately 2 to 3 minutes. Do not allow the vial to immerse near the cap line seal while thawing.
2. Once thawed, wipe the outside of the vial with 70% ethanol before opening. Using a 1 mL syringe equipped with a 27 gauge 1/2 inch needle, remove approximately 200 µL to 300 µL from the vial.
3. Wipe the injection site of the mouse with 70% ethanol and inject the sample intraperitoneally at 50 µL to 100 µL per mouse (approximately 3 mice for most applications).

### Monitoring parasitemia:

1. Starting 3 days post-inoculation, monitor the growth of parasites by tail vein bleed sampling and Giemsa-stained thin blood smear microscopy at 1- to 2-day intervals.
2. Passage the strain when the infection is at or near the first peak of parasitemia (> 5%). This will normally occur within one week of inoculation.

$$\% \text{ parasitemia} = (\text{Infected RBC} / \text{Total RBC}) \times 100$$

Note: Do not directly inject freshly thawed parasites from cryopreserved stocks by the intravenous (IV) route, as these samples contain cryoprotectant, anticoagulant and may contain traces of lysed or coagulated red blood cells. Direct IV inoculation from cryopreserved stock may result in pulmonary embolism or shock in mice.

### Passaging:

1. Anesthetize infected mice by CO<sub>2</sub>/O<sub>2</sub> inhalation. Collect the blood by orbital bleeding or from the tail vein into 25 mL of cold 1× PBS-heparin anticoagulant solution (please refer to Appendix I for preparation instructions).
2. Inject the sample into each of the uninfected mice (approximately 10 mice) as described in Inoculation step #3 above.
3. Monitor parasitemia as described above and passage as needed.

### Citation:

Acknowledgment for publications should read "The following reagent was obtained through BEI Resources, NIAID, NIH: Plasmodium berghei, Strain (ANKA) 507m6cl1, MRA-867, contributed by Chris Janse and Andrew Waters."

### Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the following publication: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, and National Institutes of Health. Biosafety in Microbiological and Biomedical Laboratories. 6th ed. Washington, DC: U.S. Government Printing Office, 2020; see [www.cdc.gov/biosafety/publications/bmbl5/index.htm](https://www.cdc.gov/biosafety/publications/bmbl5/index.htm).

All blood cultures should be handled with appropriate safety precautions necessary for the handling of bloodborne pathogens. Personnel must be trained in accordance with their institutional policy regarding bloodborne pathogens.

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### References:

1. Janse, C. J. and A. P. Waters. Personal Communication.
2. Janse, et al. "High Efficiency Transfection of *Plasmodium berghei* Facilitates Novel Selection Procedures." *Mol. Biochem. Parasitol.* 145 (2006): 60-70. PubMed: 16242190.
3. Ramiro, R. S., S. E. Reece and D. J. Obbard. "Molecular Evolution and Phylogenetics of Rodent Malaria Parasites." *BMC Evol. Biol.* 12 (2012): 219. PubMed: 23151308.
4. Hall, N., et al. "A Comprehensive Survey of the *Plasmodium* Life Cycle by Genomic, Transcriptomic, and Proteomic Analyses." *Science* 307 (2005): 82-86. PubMed: 15637271.
5. Otto, T. D., et al. "A Comprehensive Evaluation of Rodent Malaria Parasite Genomes and Gene Expression." *BMC Biol.* 12 (2014): 86. PubMed: 25359557.
6. Peters, W. and B. L. Robinson (1999), "Chapter 92 -- Malaria." In *Handbook of Animal Models of Infection*. Eds. O. Zak and M. Sande, Academic Press: London, pp. 757-773.

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## APPENDIX I: CRYOPRESERVATION

**Note:** Only immature blood-stage parasites (rings) are viable by this method. Parasitemia of 1% or higher of ring-stage parasites is recommended for cryopreservation. All steps should be carried out in a biosafety cabinet under proper air flow.

1. Prepare a 1× PBS-heparin anticoagulant solution using sterile 1× PBS (pH ~ 7.2) without calcium or magnesium (ATCC® 30-2200™) adjusted to contain 30 Units/mL sterile heparin.
2. Harvest parasitized mouse blood into 25 × volume ice cold sterile 1× PBS-heparin anticoagulant solution and place on ice.
3. Centrifuge the diluted blood culture at 1000 × g for 5 minutes at 4°C.
4. Aspirate the supernatant carefully. Measure the volume of packed red blood cells using centrifuge tube graduations or standard volume controls.
5. To the volume of packed red blood cells, add dropwise one volume of cold (4°C) Glycerolyte 57 solution (Fenwal, Cat. No. 4A7831, or equivalent). Let stand for 5 minutes at room temperature.
6. Add dropwise an additional volume of cold Glycerolyte 57 solution to the pellet. Mix well and aliquot 0.5 mL into 1.5 mL sterile cryopreservation vials.
7. Place the vials in a controlled-rate freezing unit. From room temperature, cool the vials at -1°C/min to -40°C. If the freezing unit can compensate for the heat of fusion, maintain rate at -1°C/min through this phase. At -40°C, plunge vials into liquid nitrogen. Alternatively, place the vials in a Nalgene 1°C freezing container. Place the container at -80°C for 24 to 48 hours and then plunge vials into liquid nitrogen.
8. Store in either the vapor or liquid phase of a nitrogen refrigerator (-130°C or colder).