SUPPORTING INFECTIOUS DISEASE RESEARCH

CapSM-K-SV40 Bat (*Carollia perspicillata*) Immortalized Cell Line

Catalog No. NR-59778

For research use only. Not for use in humans.

Contributor:

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Manufacturer:

BEI Resources

Product Description:

CaPSM-K-SV40 is an immortalized bat cell line derived from the kidney of a healthy three-year-old male Seba's short-tailed bat (*Carollia perspicillata*) from a disease-free bat colony. The primary cells were screened with a panel of pseudotyped virus-like particles containing glycoproteins from a wide range of viruses and selected for cells that were most susceptible to entry from the different virus glycoproteins. The selected cell lines were immortalized by transduction with SV40 T-antigen.

Note: The recommended cell life expectancy is 30 passages.

CaPSM-K-SV40 is a stable and accessible cell line, that has been tested for immortalization, susceptibility to virus infection, and response to immune stimulation and is suitable for batderived *in vitro* cellular model studies. These kidney-derived cell lines are susceptible to a range of viruses, including Middle Eastern respiratory syndrome coronavirus (MERS-CoV), vesicular stomatitis virus (VSV), and ortho hantaviruses. The immuno-competency of the cells expressing the interferon beta gene was tested with interferon pathway stimulation experiments and demonstrated an intact innate immune response.¹

Material Provided:

Each vial contains approximately 1 mL of cell culture suspension frozen in cell growth medium (90%) and DMSO (10%) cryopreservative. Sufficient cells are provided to initiate at least one new culture. The cell count, expressed as cells/vial, is shown on individual certificates of analysis for each lot.

Packaging/Storage:

This product was packaged aseptically, in cryovials. It should be stored at -100°C or colder, preferably in the vapor phase of a liquid nitrogen freezer. Storage at -60°C will result in loss of viability. To ensure the highest level of viability, the vial should be thawed and the culture initiated as soon as possible upon receipt. Any warming of the product during shipping and transfer must be avoided, as this will adversely affect the viability of the product after thawing. For the transfer between freezers and shipping, the cells may be placed on dry ice for brief periods, although the use of a portable liquid nitrogen carrier is preferred. Please read the following recommendations prior to reconstituting this material.

Safety Precautions:

When handling frozen vials, it is highly recommended that protective gloves, a lab coat, and a full-face mask be worn. Even brief exposure to the ultra-cold temperature can cause tissue damage from frostbite. Also, some vials may slowly fill with liquid nitrogen if they have been immersed during cryogenic storage. When thawing, the liquid nitrogen may rapidly expand as it changes to gas, breaking the vial or cap with explosive force and sending debris flying with enough velocity to cause injury. Store and use in areas with adequate ventilation.

Thawing and Growth:

Prior to thawing the bat cells, prepare growth medium (GM) for use. Bat cells are grown in Dulbecco's Modified Eagle's Medium (ATCC[®] 30-2002) containing 4 mM L-glutamine, 4500 mg/L-glucose, 1 mM sodium pyruvate, and 1500 mg/mL sodium bicarbonate, supplemented with 10% fetal bovine serum (ATCC[®] 30-2020). This GM is formulated for use with a 5% CO₂ in the air atmosphere.

Rapidly thaw the vial of bat cells in a 37° C water bath with gentle agitation. To reduce the risk of contamination, keep the cap and O-ring of the vial out of the water and repeatedly check the cap for tightness during thawing. Remove from the water bath immediately when thawed. Dry the vial with a sterile wiper, decontaminate using a wiper soaked with 70% isopropyl alcohol, and let the vial air dry. Aseptically open the vial, remove the vial contents, and add 4 mL of GM in a centrifuge tube. Centrifuge the cell suspension at 125 to 200 × g for 8 to 10 minutes at 18 to 25° C. Discard the supernatant and resuspend the cell pellet in 10 mL of prewarmed GM. Transfer the cell suspension into a 75 cm² tissue culture flask. Incubate the new culture at 37° C and 5% CO₂. Replace the GM with fresh GM every 2 to 3 days and incubate until the cell sheet is approximately 80% confluent.

Sub-culture procedure:

Aseptically remove the GM and discard. Briefly rinse the cell layer with 4 to 15 mL of Ca²⁺- and Mg²⁺-free Dulbecco's phosphate-buffered saline (PBS) to remove all traces of serum. Discard the PBS. Add 2 to 8 mL of 0.05% trypsin-EDTA to the culture flask and incubate the flask at 37°C until cell layer is dispersed (usually within 3 minutes but no longer than 15 minutes). *Note: To avoid clumping, do not agitate the cells by hitting or shaking the flask.* Add an equal volume of GM and aspirate cells by gently pipetting. Perform a cell count and add appropriate aliquots of the cell suspension to new culture vessels at a sub-cultivation ratio of 1:3 to 1:4. Adjust the volume of GM to 15 to 20 mL for a 75 cm² flask. Incubate cultures at 37°C and 5% CO₂. Replace the GM with fresh GM every 2 to 3 days and incubate until the cell sheet is approximately 80% confluent.

Citation:

Acknowledgment for publications should read "The following reagent was obtained through BEI Resources, NIAID, NIH:

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CapSM-K-SV40 Bat (*Carollia perspicillata*) Immortalized Cell Line, NR-59778."

Biosafety Level: 2

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the following publication: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, and National Institutes of Health. Biosafety in Microbiological and Biomedical Laboratories (BMBL). Current Edition. Washington, DC: U.S. Government Printing Office.

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References:

 Letko, M., et al. "Viral Susceptibility and Innate Immunity Competency of *Carollia perspicillata* Bat Cells Produced for Virological Studies." <u>bioRxiv</u> Preprint. 2024 Nov 19:2024.11.19.624190. PubMed: 39605657.

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