

**Toxoplasma gondii, Strain RH MIC2-GLuc-C-myc**

**Catalog No. NR-51148**

**Product Description:** *Toxoplasma gondii* (*T. gondii*), strain RH MIC2-GLuc-C-myc was deposited to BEI Resources as a transgenic strain derived from the virulent Type I strain RH, created by co-transfection with pMIC2-GLuc-C-myc and pBS-TUB1CatSAG1 and selected with chloramphenicol.

**Lot<sup>1</sup>: 70013168**

**Manufacturing Date: 12MAR2018**

| TEST   | SPECIFICATIONS   | RESULTS  |
|--|--|--|
| <b>Cell Morphology<sup>2</sup></b>   | Report results   | Crescent-shaped and refractile   |
| <b>Genotypic Analysis<sup>3</sup></b><br>Sequencing of 850 locus (~ 710 base pairs)<br><br>Sequencing of <i>Gaussia princeps</i> luciferase (GLuc) gene (~ 510 base pairs)   | ≥ 99% sequence identity to <i>T. gondii</i> , strain RH (GenBank: LLKL01000174.1)<br><br>≥ 99% sequence identity to synthetic construct GLuc (GenBank: MF882921.1) | 100% sequence identity to <i>T. gondii</i> , strain RH (GenBank: LLKL01000174.1) (Figure 1)<br><br>100% sequence identity to synthetic construct GLuc (GenBank: MF882921.1) (Figure 2) |
| <b>PCR Assay of Extracted DNA<sup>3</sup></b><br>850 locus <sup>4</sup><br>GLuc <sup>5</sup>   | ~ 770 base pair amplicon<br>~ 530 base pair amplicon   | ~ 770 base pair amplicon<br>~ 530 base pair amplicon   |
| <b>Phenotypic Analysis</b><br>C-myc immunofluorescence assay <sup>6</sup>  | Positive   | Positive (Figure 3)  |
| <b>Viable Cell Count by Hemacytometry<sup>3</sup></b>  | > 10 <sup>6</sup> cells per mL   | 1.4 x 10 <sup>8</sup> cells per mL   |
| <b>Viability (post-freeze)<sup>2,7</sup></b>   | Viable parasites   | Viable parasites   |
| <b>Sterility (21-day incubation)<sup>2</sup></b><br>Harpo's HTYE broth <sup>8</sup> , 37°C and 26°C, aerobic<br>Trypticase soy broth, 37°C and 26°C, aerobic<br>Sabouraud broth, 37°C and 26°C, aerobic<br>DMEM with 10% FBS, 37°C, aerobic<br>Sheep blood agar, 37°C, aerobic<br>Sheep blood agar, 37°C, anaerobic<br>Thioglycollate broth, 37°C, anaerobic | No growth<br>No growth<br>No growth<br>No growth<br>No growth<br>No growth<br>No growth  | No growth<br>No growth<br>No growth<br>No growth<br>No growth<br>No growth<br>No growth  |
| <b>Mycoplasma Contamination<sup>2</sup></b><br>DNA Detection by PCR  | None detected  | None detected  |

<sup>1</sup>NR-51148 was produced by cultivation of the deposited material in human foreskin fibroblast cells (ATCC<sup>®</sup> CRL-1634<sup>™</sup>) with cell cultivation medium for parasites (ATCC<sup>®</sup> medium 2222: adjusted to contain 10% heat-inactivated fetal bovine serum). The culture was propagated for 3 days at 37°C in an aerobic atmosphere with 5% CO<sub>2</sub> until lysis of the host cell monolayer was reached.

<sup>2</sup>Testing completed on vial, post-freeze material.

<sup>3</sup>Testing completed on bulk material prior to vialing and freezing.

<sup>4</sup>Primer sequences and PCR conditions are available upon request.

<sup>5</sup>Primer sequences as reported in Brown, K. M., S. Lourido and L. D. Sibley. "Serum Albumin Stimulates Protein Kinase G-dependent Microneme Secretion in *Toxoplasma gondii*." *J. Biol. Chem.* 291 (2016): 9554-9565. PubMed: 26933037.

<sup>6</sup>Immunolabeling of *T. gondii*, strain RH MIC2-GLuc-C-myc was observed using a C-myc tag (Myc.A7) mouse monoclonal antibody (ThermoFisher, MA1-21316; 1:1000 dilution), followed by fluorescein isothiocyanate (FITC)-conjugated goat anti-mouse IgG (Invitrogen<sup>®</sup> A16073; 1:2000 dilution) as secondary antibody (Figure 3A) and overlaid with the 4',6-diamidino-2-phenylindole (DAPI)-stained image (Figure 3B).

<sup>7</sup>Viable cells and signs of infection were seen after 4 days under cultivation conditions at 37°C.

<sup>8</sup>Atlas, Ronald M. *Handbook of Microbiological Media*. 3rd ed. Ed. Lawrence C. Parks. Boca Raton: CRC Press, 2004, p. 798.

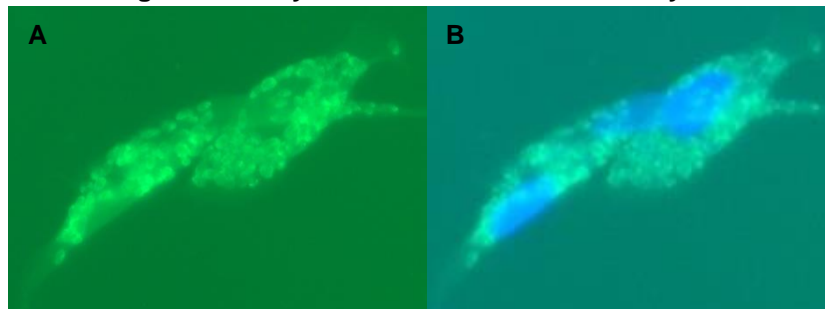
**Figure 1: 850 Locus Amplicon Sequence**

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CCTCGTCCAG CCGATGCTGC ATGGCTGCCA CCCCTTCCTC GTAGCCCCC TGTCGGTGAG GCAACTGGTC
CCCGTGGGGT CTTTAAAAGG CTCAACCGGC TACCGGGGCT ACGCGAAGGC GACCCCTCTC CACGAGAAGG
CCCTCCGACC TTATCGACGC CCGTTCGCCC TGGCGACGGC TTGCCATCGC TTCTGGGTGT CGGCGCTGCT
TTCCCTGGAG GCATCCCTGT TTGTGGGGAG GAAGCAGTAG TGGCACTAAT GGGTGCCTGT GCCTGCCTCC
CTCCTTGCTC CGGTGAGCTG CTGCCCCCA CAGGTCCCTC TTGCTGCATG CCTCGTGCAG GAGCGCTTGG
CGTCGGTGAG TCACCATCTG ACGGTGAAAC TGAGCTGTCG CCCAAGCCGC TGCTGCCTGA CGACGAAGAA
GGCCACCTG AGTGGATTAT GACGACGTC A CCTCTGGCC CGCCCTCAGA GCCACGAAA AATGAAGCAA
GACGCGGGCC TTGCAGTGGG GACGGTGATG GCGGCGAACG TTTTCCGGGA ACGTGTGTTT CGATGTCGCT
TTTCGGGGAT GTTTCCTTTG GAAGCGAGCC CTTTGCGCCG CAGCACGGAC TTTGTGTGTC TGCTGGTACA
CGGACAGTCT CTGAAGGACT CCCCCTTGCA GGTGTGGAAT GTAAAGGCCG CTCTACGGGT TTCCCCCAG
ATGCTGGAGA AGG
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**Figure 2: GLuc Gene Amplicon Sequence**

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GAAGCCACC GAGAACAACG AAGACTTCAA CATCGTGGCC GTGGCCAGCA ACTTCGCGAC CACGGATCTC
GATGCTGACC GCGGGAAGTT GCCCGCAAG AAGCTGCCGC TGGAGGTGCT CAAAGAGATG GAAGCCAATG
CCCGAAAAGC TGGCTGCACC AGGGGCTGTC TGATCTGCCT GTCCACATC AAGTGCACGC CCAAGATGAA
GAAGTTCATC CCAGGACGCT GCCACACCTA CGAAGGCGAC AAAGAGTCCG CACAGGGCGG CATAGGCGAG
GCGATCGTCG ACATTCTGA GATTCTGGG TTCAAGGACT TGGAGCCCAT GGAGCAGTTC ATCGCACAGG
TCGATCTGTG TGTGGACTGC ACAACTGGCT GCCTCAAAGG GCTTGCCAAC GTGCAGTGTG CTGACCTGCT
CAAGAAAGTGG CTGCCGCAAC GCTGTGCGAC CTTTGCCAGC AAGATCCAGG GCCAGGTGGA CAAGATCAAG
GGGGCCGGTG GTGACC
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**Figure 3: C-myc Immunofluorescence Assay**



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Program Manager or designee, ATCC Federal Solutions

27 AUG 2018

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