

Certificate of Analysis for NR-59622

Center for Biologics Evaluation and Research Virus Next Generation Sequencing Reference Reagents

Catalog No. NR-59622

Product Description:

NR-59622 consists of a 5-member panel of viral preparations identified by the World Health Organization (WHO) as standards for high-throughput sequencing. These preparations are intended for next generation sequencing (NGS) studies for method establishment, verification, qualification and validation. Viruses should be used directly and not grown or amplified before use.

Lot: 70064122 Assembly Date: OCT 2023

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Table 1: Kit Components

| COMPONENT NUMBER | DESCRIPTION | HOST CELL LINE | LOT NUMBER | MANUFACTURING DATE |
|---------------------|--|---|---------------|-----------------------|
| SC-VR-6000P™ | Custom preparation of porcine circovirus type 1 | PK(15) porcine kidney cells (ATCC® CCL-33™) | 63856605 | 08DEC2015 |
| SC-VR-6001P™ | Custom preparation of mammalian orthoreovirus type 1, strain Lang | LLC-MK2 derivative Rhesus monkey kidney cells (ATCC® CCL-7.1™) | 63633442 | 28JUL2015 |
| SC-VR-6002P™ | Custom preparation of feline leukemia virus, strain Thielen | FL74-UCD-1 cat lymphoblast cells (ATCC [®] CRL-8012™) | 63856597 | 18APR2016 |
| SC-VR-6003P™ | Custom preparation of human respiratory syncytial virus, strain A2 | HEp-2 cells (ATCC [®] CCL-23™) | 63633439 | 14JUL2015 |
| SC-VR-6004P™ | Custom preparation of Epstein-Barr virus (HHV-4), strain B95-8 | B95-8 Leukocyte Marmoset culture (ATCC [®] CRL-1612™) | 63633440 | 03SEP2015 |

Table 2: Custom preparation of porcine circovirus type 1 (SC-VR-6000P™)^{1,2,3}

| table 2. Gustom preparation of poreme encovirus type 1 (00-411-00001) | | | |
|---|--|--|--|
| Test / Method | Specification | Result | |
| Titer (Post-vial) ^{4,5} | ≥ 1 × 10 ⁶ TCID ₅₀ /mL | 1.2 × 10 ⁷ TCID ₅₀ /mL | |
| Genome Copy Number by ddPCR (Post-vial) ^{5,6,7} | ≥ 1 × 10 ¹⁰ genome copies/mL | 2.7 × 10 ¹¹ genome copies/mL | |
| Test for Mycoplasma Contamination DNA detection by PCR of test article nucleic acid [Universal Mycoplasma Detection Kit (ATCC® 30-1012K™)] | None detected | None detected | |
| Sterility Test (BacT/ALERT 3D) iAST bottle (aerobic) at 32°C, 14-day incubation iNST bottle (anaerobic) at 32°C, 14-day incubation | No growth No growth | No growth No growth | |

¹Porcine circovirus type 1 (PCV1) was grown in PK(15) porcine kidney cells (ATCC[®] CCL-33™) at 37°C with 5% CO₂. PK(15) cells are known to contain porcine endogenous retrovirus [Pol. J. Microbiol. (2012), 61: 211-215. PubMed: 29334069].

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²Preparation was vialed in 10 mM Tris-HCl, 135 mM NaCl, 0.5% BSA and 5% trehalose and may contain residual cellular nucleic acids.

³Stability testing of the genome copy number and titer completed in 2018 and 2022 shows that the material maintains these characteristics when stored at -80°C.

⁴16 days in ST cells (ATCC® CRL-1746™) at 37°C with 5% CO₂, as determined by endpoint PCR with PCV1 specific primers.

⁵Test result from April 2018.



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⁶ddPCR data was obtained post-vial from 9 replicates on the BioRad QX200 Droplet Digital PCR (ddPCR™) System; gene assayed was replication associated protein.

⁷ddPCR samples contain virus genomes and may have residual mRNAs.

Table 3: Custom preparation of mammalian orthoreovirus type 1, strain Lang (SC-VR-6001P™)^{1,2,3}

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|--|--|---|--|
| Test / Method | Specification | Result | |
| Titer (Post-vial) ^{4,5} | ≥ 1 × 10 ⁶ TCID ₅₀ /mL | 1.1 × 10 ¹⁰ TCID ₅₀ /mL | |
| Genome Copy Number by ddPCR (Post-vial) ^{5,6,7} | ≥ 1 × 10 ¹⁰ genome copies/mL | 1.4 × 10 ¹⁰ genome copies/mL | |
| Test for Mycoplasma Contamination | | | |
| DNA detection by PCR of test article nucleic acid [Universal Mycoplasma Detection Kit (ATCC® 30-1012K™)] | None detected | None detected | |
| Sterility Test (BacT/ALERT 3D) | | | |
| iAST bottle (aerobic) at 32°C, 14-day incubation iNST bottle (anaerobic) at 32°C, 14-day incubation | No growth | No growth | |
| 1140 F bottle (arraerobic) at 52 G, 14-day incubation | No growth | No growth | |

¹Mammalian orthoreovirus (MRV) type 1, strain Lang, was grown in LLC-MK2 derivative Rhesus monkey kidney cells (ATCC® CCL-7.1™) at 37°C with 5% CO₂ and humidity.

Table 4: Custom preparation of feline leukemia virus, strain Thielen (SC-VR-6002P™)^{1,2,3}

| Table 4. Ouston preparation of femic leakenna virus, strain Thicien (00-viv-0002) | | | |
|---|--|--|--|
| Test / Method | Specification | Result | |
| Titer (Post-vial) ^{4,5} | ≥ 1 × 10 ⁶ TCID ₅₀ /mL | 2.3 × 10 ⁷ TCID ₅₀ /mL | |
| Genome Copy Number by ddPCR (Post-vial) ^{5,6,7} | ≥ 1 × 10 ¹⁰ genome copies/mL | 5.3×10^{10} genome copies/mL | |
| Test for Mycoplasma Contamination DNA detection by PCR of test article nucleic acid [Universal Mycoplasma Detection Kit (ATCC® 30-1012K™)] | None detected | None detected | |
| Sterility Test (BacT/ALERT 3D) iAST bottle (aerobic) at 32°C, 14-day incubation iNST bottle (anaerobic) at 32°C, 14-day incubation | No growth No growth | No growth No growth | |

¹Feline leukemia virus (FLV), strain Thielen, was grown in FL74-UCD-1 cat lymphoblast cells (ATCC[®] CRL-8012™) at 36°C.

Table 5: Custom preparation of human respiratory syncytial virus, strain A2 (SC-VR-6003P™)^{1,2,3}

| Test / Method | Specification | Result |
|--|--|--|
| Titer (Post-vial) ^{4,5} | ≥ 1 × 10 ⁶ TCID ₅₀ /mL | 1.1 × 10 ⁶ TCID ₅₀ /mL |
| Genome Copy Number by ddPCR (Post-vial) ^{5,6,7,8} | ≥ 1 × 10 ¹⁰ genome copies/mL | 1.0 × 10 ⁹ genome copies/mL |

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²Preparation was vialed in 10 mM Tris-HCl, 135 mM NaCl, 0.5% BSA and 5% trehalose and may contain residual cellular DNA.

³Stability testing of the genome copy number and titer completed in 2018 and 2022 shows that the material maintains these characteristics when stored at -80°C.

⁴9 days on LLC-MK2 cells (ATCC[®] CCL-7.1™) at 37°C with 5% CO₂ and humidity, as determined by CPE.

⁵Test result from April 2018.

⁶ddPCR data was obtained post-vial from 9 replicates on the BioRad QX200 Droplet Digital PCR (ddPCR™) System; gene assayed was L2. ⁷ddPCR samples contain virus genomes and may have residual mRNAs.

²Preparation was vialed in 10 mM Tris-HCl, 135 mM NaCl, 0.5% BSA and 5% trehalose and may contain residual cellular nucleic acids.

³Stability testing of the genome copy number and titer completed in 2018 and 2022 shows that the material maintains these characteristics when stored at -80°C.

⁴7 days in MYA-1 cells (ATCC[®] CRL-2417[™]) at 37°C with 5% CO₂ and humidity, as determined by endpoint PCR with FLV specific primers. ⁵Test result from August 2018.

⁶ddPCR data was obtained post-vial from 9 replicates on the BioRad QX200 Droplet Digital PCR (ddPCR™) System; gene assayed was protease. ⁷ddPCR samples contain virus genomes and may have residual mRNAs.



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| Test / Method | Specification | Result |
|---|---------------|---------------|
| Test for Mycoplasma Contamination | | |
| DNA detection by PCR of test article nucleic acid [Universal Mycoplasma Detection Kit (ATCC [®] 30-1012K [™])] | None detected | None detected |
| Sterility Test (BacT/ALERT 3D) | | |
| iAST bottle (aerobic) at 32°C, 14-day incubation | No growth | No growth |
| iNST bottle (anaerobic) at 32°C, 14-day incubation | No growth | No growth |

¹Human respiratory syncytial virus (hRSV), strain A2, was grown in HEp-2 cells (ATCC[®] CCL-23[™]) at 37°C with 5% CO₂ and humidity.

Table 6: Custom preparation of Epstein-Barr virus (HHV-4), strain B95-8 (SC-VR-6004P™)^{1,2,3}

| Table 6. Custom preparation of Epstein-Barr virus (HHV-4), strain B95-8 (SC-VK-6004F) | | | |
|---|--|--|--|
| Test / Method | Specification | Result | |
| Titer (Post-vial) ^{4,5} | ≥ 1 × 10 ⁶ TCID ₅₀ /mL | 1.1 × 10 ⁷ TCID ₅₀ /mL | |
| Genome Copy Number by ddPCR (Post-vial) ^{5,6,7,8} | ≥ 1 × 10 ¹⁰ genome copies/mL | 3.7 × 10 ⁸ genome copies/mL | |
| Test for Mycoplasma Contamination DNA detection by PCR of test article nucleic acid [Universal Mycoplasma Detection Kit (ATCC® 30-1012K™)] | None detected | None detected | |
| Sterility Test (BacT/ALERT 3D) iAST bottle (aerobic) at 32°C, 14-day incubation iNST bottle (anaerobic) at 32°C, 14-day incubation | No growth No growth | No growth No growth | |

¹Epstein-Barr virus [human herpes virus 4 (HHV-4)], strain B95-8, was isolated from B95-8 Leukocyte Marmoset culture (ATCC® CRL-1612™) grown at 37°C with humidity with 5% CO₂. The B95-8 marmoset cell line is known to contain squirrel monkey retrovirus [Virology. (1995), 209: 374-383.

/Sonia Bjorum Brower/ Sonia Bjorum Brower

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Technical Manager or designee, ATCC Federal Solutions

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²Preparation was vialed in 10 mM Tris-HCl, 135 mM NaCl, 0.5% BSA and 5% trehalose and may contain residual cellular nucleic acids.

³Stability testing of the genome copy number and titer completed in 2018 and 2022 shows that the material maintains these characteristics when stored at -80°C.

⁴⁸ days in HEp-2 cells (ATCC[®] CCL-23[™]) at 37°C with 5% CO₂ and humidity, as determined by Immunofluorescence Light Diagnostics [™] Respiratory Syncytial Virus FITC Reagent (Millipore catalog # 5022).

⁵Test result from April 2018.

⁶ddPCR data was obtained post-vial from 9 replicates on the BioRad QX200 Droplet Digital PCR (ddPCR™) System; gene assayed was N protein. ⁷ddPCR samples contain virus genomes and may have residual mRNAs.

⁸The genome copy number for hRSV, strain A2 is below the current specifications but does not negatively impact the final product.

²Preparation was vialed in 10 mM Tris-HCl, 135 mM NaCl, 0.5% BSA and 5% trehalose and may contain residual cellular nucleic acids.

³Stability testing of the genome copy number and titer completed in 2018 and 2022 shows that the material maintains these characteristics when stored at -80°C.

⁴60 days in irradiated human lung fibroblast cells (ATCC[®] 55-X™) at 37°C with 5% CO₂ and humidity, as determined by transformation.

⁵Test result from May/June 2018.

⁶ddPCR data was obtained post-vial from 9 replicates on the BioRad QX200 Droplet Digital PCR (ddPCR™) System; gene assayed was EBER1 noncoding RNA.

⁷ddPCR samples contain virus genomes and may have residual mRNAs.

⁸The genome copy number for HHV-4, strain B95-8 is below the current specifications but does not negatively impact the final product.