

Pan-Rickettsia Quantitative PCR (qPCR) Assay Detection Kit

Catalog No. NR-35520

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Contributor and Manufacturer:

BEI Resources

Product Description:

The Pan-Rickettsia Quantitative PCR Assay Detection Kit (NR-35520) is designed to detect and quantitate the presence of *Rickettsia* species. The assay consists of the following components:

- 1) Forward and reverse primers (NR-35354 and NR-35355, respectively)
- 2) Probe designed with 6-carboxyfluorescein (6-FAM) at the 5' end and both a minor groove binding moiety (MGB) and a non-fluorescent quenching dye (NFQ) at the 3' end (NR-35356)
- 3) Plasmid-based standard containing *Rickettsia prowazekii* ribosomal RNA (rRNA) gene sequences (NR-35519)

The plasmid-based standard, NR-35519, is linearized pIDTBlue containing a one kilobase synthetic DNA insert corresponding to the 16S rRNA gene of the Madrid E strain of *Rickettsia prowazekii*. A ten nucleotide sequence within the 135 base pair target amplicon is inverted to facilitate discrimination between the presence of authentic *Rickettsia* genetic material and false positives resulting from plasmid contamination. NR-35520 is expected to detect virtually all *Rickettsia* species owing to the high degree of conservation of the 16S rRNA gene within the genus, and of the primer and probe sequences in particular.

Each kit contains enough primer and probe for approximately 250 reactions using the assay protocol outlined in Appendix I. Lot-specific Certificates of Analysis for individual components are available upon request.

Material Provided:

Each vial of primer contains 200 µL in 0.2 mM Tris, 0.02 mM EDTA, pH 8.0. Each vial of probe contains 200 µL in 0.5 mM Tris, 0.05 mM EDTA, pH 8.0. Each vial of plasmid-based standard contains 2 × 10¹⁰ molecules in 100 µL nuclease-free water. A representative assay is shown in Appendix II.

Packaging/Storage:

Primers, probe, and standard were packaged aseptically in screw-capped plastic cryovials. The product is provided frozen on dry ice and should be stored at -20° C or colder upon arrival. Freeze-thaw cycles should be minimized. Probe samples should be kept in the dark at all times.

Citation:

Acknowledgment for publications should read “The following reagent was obtained through BEI Resources, NIAID, NIH: Pan-Rickettsia Quantitative PCR (qPCR) Assay Detection Kit, NR-35520.”

Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the following publication: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, and National Institutes of Health. Biosafety in Microbiological and Biomedical Laboratories. 5th ed. Washington, DC: U.S. Government Printing Office, 2009; see www.cdc.gov/biosafety/publications/bmbl5/index.htm.

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APPENDIX I

Quantitative PCR Assay for the Detection and Quantitation of *Rickettsia* Species

Recommended Reagents/Equipment

Reagent/Equipment	Source	Catalog #
Pan-Rickettsia Quantitative PCR Probe	BEI Resources	NR-35356
Pan-Rickettsia Quantitative PCR Forward Primer	BEI Resources	NR-35354
Pan-Rickettsia Quantitative PCR Reverse Primer	BEI Resources	NR-35355
Plasmid Containing Pan-Rickettsia B9R Gene Sequences, Linearized	BEI Resources	NR-35519
iTaq™ DNA Polymerase	BioRad	170-8870
Molecular Grade Water	ATCC®	60-2450
CFX 96-Well Plate Thermal Cycler	BioRad	184-5096

Preparation of Plasmid-Based Standard Curve Samples

Dilution Tube	Volume (µL)	Volume Molecular Grade Water (µL)	Concentration (Copies per 5 µL) ¹
Undiluted NR-35519	---	---	1 × 10 ⁹
1	5 of undiluted NR-35519	45	1 × 10 ⁸
2	5 of Tube 1	45	1 × 10 ⁷
3	5 of Tube 2	45	1 × 10 ⁶
4	5 of Tube 3	45	1 × 10 ⁵
5	5 of Tube 4	45	1 × 10 ⁴
6	5 of Tube 5	45	1000
7	5 of Tube 6	45	100
8	5 of Tube 7	45	10
9	5 of Tube 8	45	1

Reaction Mix¹

Reagent	Stock Concentration	Volume per Reaction (µL)
Molecular Grade H ₂ O	N/A	11.875
iTaq™ buffer	10X	2.5
iTaq™ DNA Polymerase	5 U per µL	0.125
dNTPs	10 mM	0.5
MgCl ₂	50 mM	3.0
Probe ^{2,3} - NR-35356	5 µM	0.5
Forward Primer ² - NR-35354	10 µM	0.75
Reverse Primer ² NR-35355	10 µM	0.75
Nucleic acid sample	N/A	5
		Total – 25 µL

¹Reaction mix should be kept on bench-top cooler until ready for use.

²Primers and probe are supplied at working stock concentrations.

³6-carboxyfluorescein probe must be protected from light at all times.

Cycling Protocol

Cycle	# of Repeats	Step	Conditions
1	1	1	95.0°C for 3 minutes
2	40	1	95.0°C for 15 seconds
		2	60.8°C for 15 seconds

Instructions

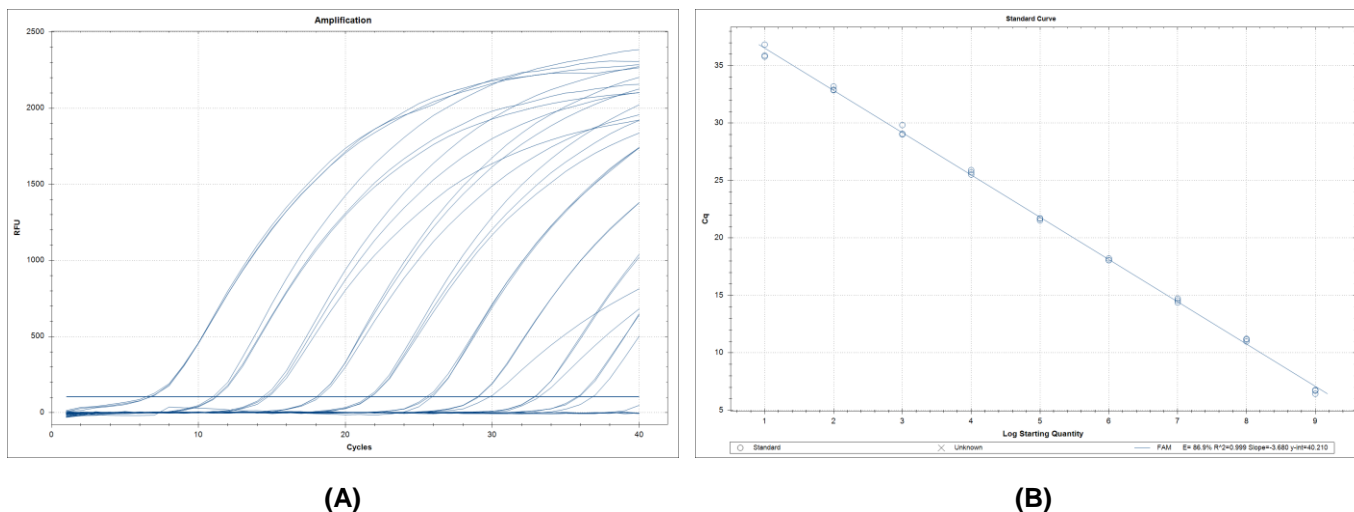
1. Prepare unknown nucleic acid samples.
2. This assay was developed using Bio-Rad reagents and detection system. Please refer to the CFX System Manual for information regarding plate and run setup.
3. When analyzing the data, especially the standard curve, it is important that the PCR efficiency fall between 80-120% and that the C_T values are separated by approximately 3.3 cycles.

**APPENDIX II
Representative Assay Results**

TEST	SPECIFICATIONS	RESULTS
Quantitative PCR – Representative Standard Curve¹ Correlation coefficient PCR efficiency Dilution separations (C _T values) Quantitative sensitivity	≥ 0.98 85 to 110% ~ 3.33 cycles Report results	0.99 86.9% ~ 3.68 cycles 100 molecules per reaction

¹See Figure 1

Figure 1



Representative quantitative PCR cycle graph (A) and associated standard curve (B) using NR-35520. The cycle threshold (C_T) was generated using regression analysis. Per-well baseline cycles have been determined automatically. The data analysis window is set at 95% of a cycle, centered at the end of the cycle.